



NEW ZEALAND'S  
BIOLOGICAL  
HERITAGE

Ngā Koiora  
Tuku Iho

National  
**SCIENCE**  
Challenges



# Next-generation biodiversity assessment for primary industries

Robert Holdaway, 14th July 2015



**Landcare Research**  
**Manaaki Whenua**

# Acknowledgements

## Next Generation Biodiversity Assessment Project

- Ian Dickie, Jamie Wood, Kate Orwin, Andreas Makiola, Alexei Drummond
- Phase 1 & Phase 2 MBIE Smart Idea funding (till September 2016)
- Funding also provided by Landcare Research, Lincoln University Bio-Protection Centre, and the Department of Conservation

## Biological Heritage National Science Challenge

- A national framework for bioheritage assessment across natural and productive landscapes (5 year project, started July 2015)
- Key researchers: Gavin Lear, Paul Gardner, Simon Bulman, Ian Dickie, Austen Ganley; Phil Wilcox; Kevin Collins
- Numerous other researchers and stakeholders who have contributed to the conception and development of the project



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MAIDMENT THEATRE



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CANTERBURY  
*Te Whare Wānanga o Waitaha*  
CHRISTCHURCH NEW ZEALAND

# Outline

Why assess biodiversity?

Environmental DNA 101

- What, how, why?

Primary industry applications

- Demonstrating environmental performance
- Improved management practices
- Biosecurity detection and monitoring

Future research directions



# Why primary industry cares about biodiversity

- Market advantage & licence to operate
  - Biodiversity foot-printing
  - Clean green image
- Sustainable practice
  - over \$500 million spent annually on fertiliser by dairy sector
  - biological farming practices can improve fertility naturally and reduce costs long-term
- Biosecurity
  - \$ billions of export earnings at risk
  - early detection key to minimising potential impacts

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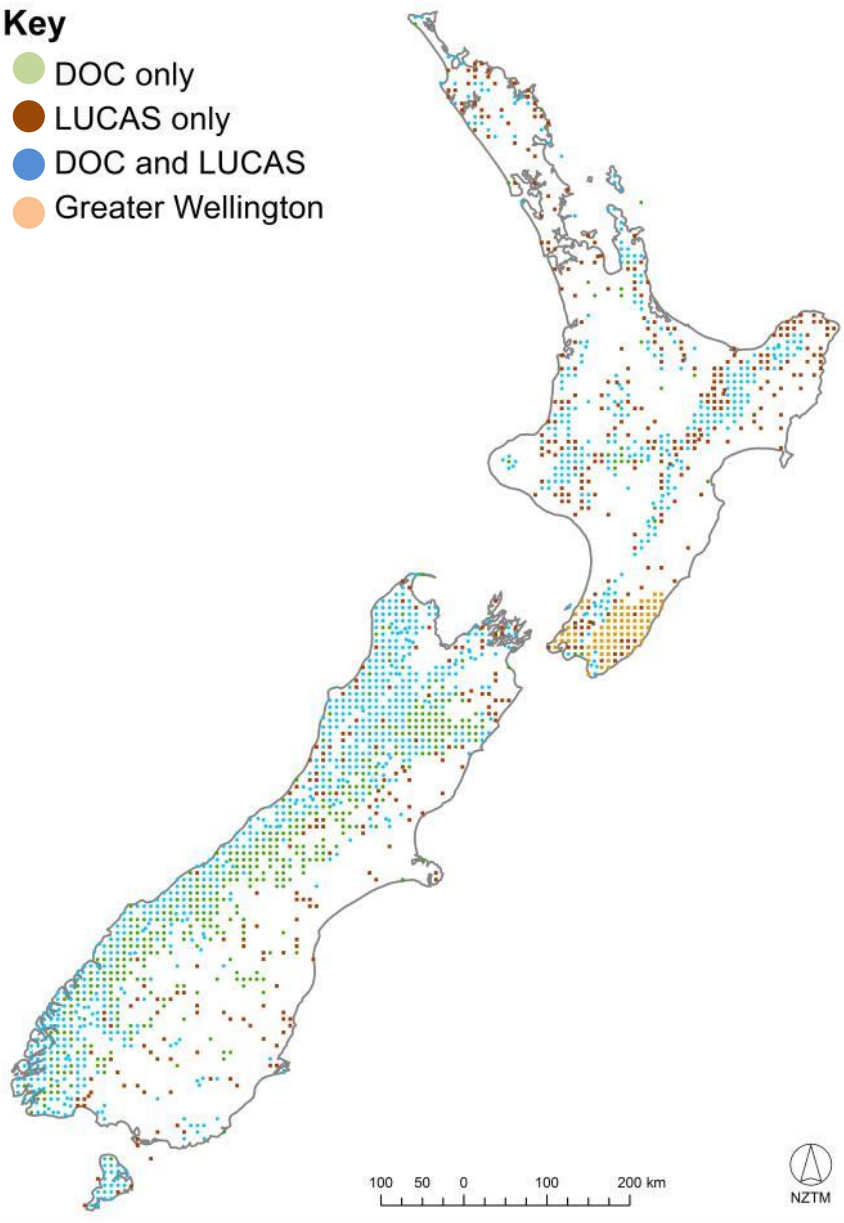




# National biodiversity assessment

## Key

- DOC only
- LUCAS only
- DOC and LUCAS
- Greater Wellington



8 km grid for State of Environment reporting

- LUCAS (MfE)
- DOC Tier 1
- Regional Councils

Harmonised sample design, methods and reporting metrics

Major limitations

- Limited primary sector coverage
- the vast majority of biodiversity is not assessed

~214 birds  
~2500 vascular plants



~20,000 insects



~22,000 fungi

Vertical scale = proportional  
to species



# Cryptic biodiversity is important for primary industry

## The good guys



**Ecosystem function**



**Agricultural productivity**

## The bad guys



**Pathogens**



**Environmental pollutants**



Next generation sequencing of environmental DNA provides a cost effective tool to measure all forms of biodiversity – especially cryptic biodiversity



# What is environmental DNA (eDNA)?

DNA extracted directly from an environmental sample



- trace DNA left behind by organisms
- undifferentiated micro-organism DNA

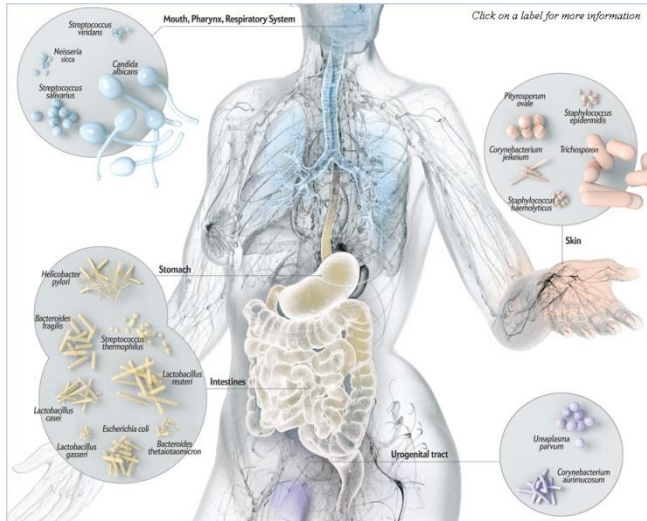


# What is Next Generation Sequencing?

- Typically used for eDNA analysis
- High throughput, massively parallel
- Commercially available service
- Massive data outputs (millions or billions of sequences per run)



# eDNA success stories



# How to assess biodiversity using eDNA?

7 fundamental steps

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7 fundamental steps

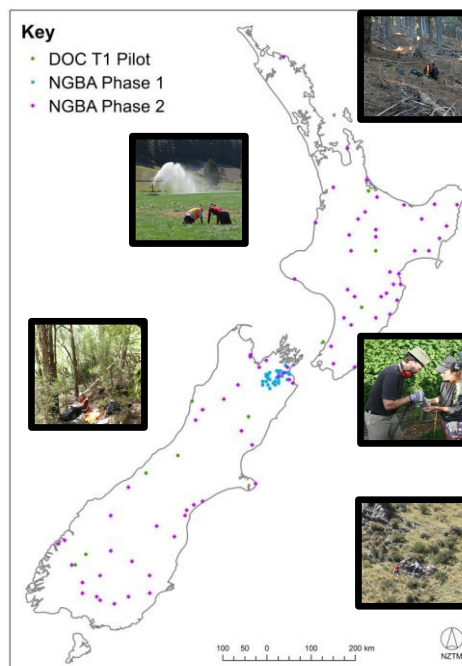
## 1. Define your question



# How to assess biodiversity using eDNA?

## 7 fundamental steps

1. Define your question
2. Design sampling





# How to assess biodiversity using eDNA?

## 7 fundamental steps

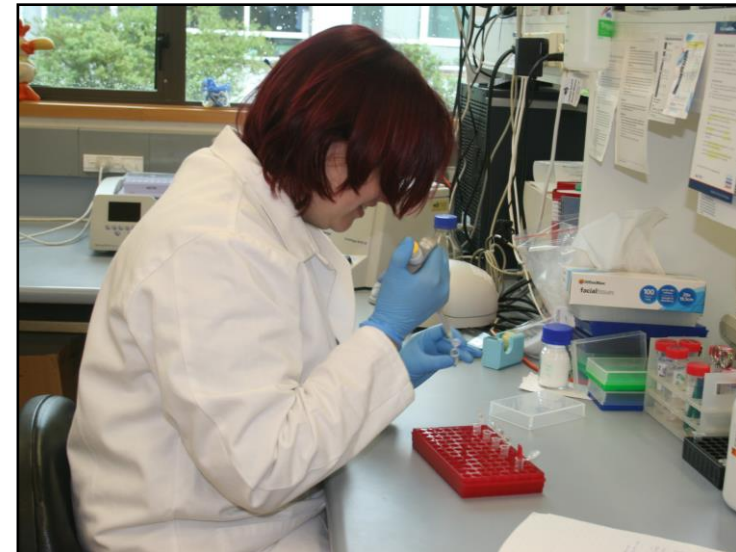
1. Define your question
2. Design sampling
- 3. Collect samples**



# How to assess biodiversity using eDNA?

## 7 fundamental steps

1. Define your question
2. Design sampling
3. Collect samples
4. **Extract DNA**



# How to assess biodiversity using eDNA?

## 7 fundamental steps

1. Define your question
2. Design sampling
3. Collect samples
4. Extract DNA
5. **Sequence DNA**

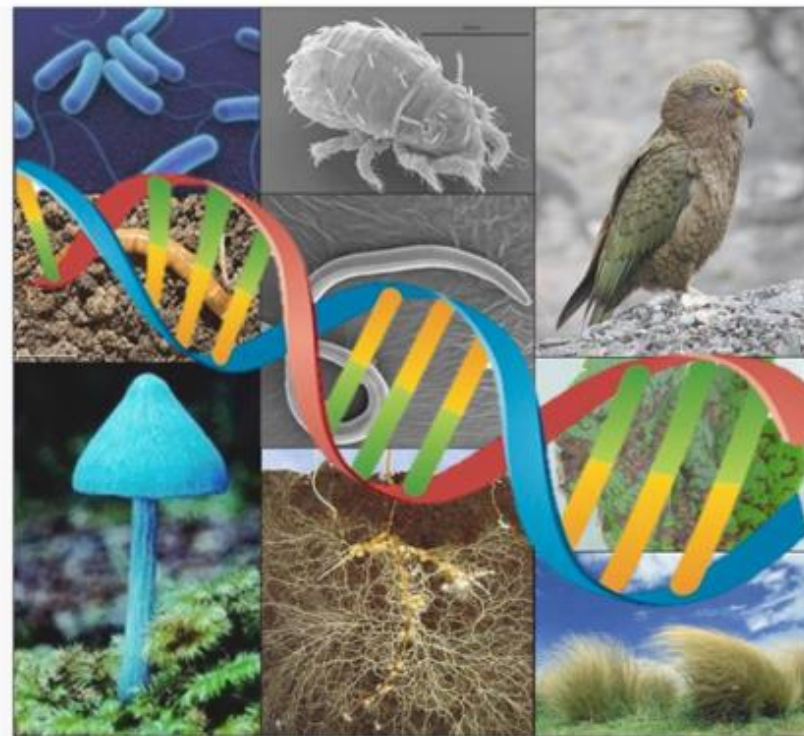
GTGAATCATCGAATCTTTGAACG  
CACATTGCGCCCCTTGGTATTCC  
GAGGGGCATGCCTATTCGAGCG  
TCATTATCACCCCTCAAGCCTAG  
CTTGGTGTTGAGACCTGCTGTC



# How to assess biodiversity using eDNA?

## 7 fundamental steps

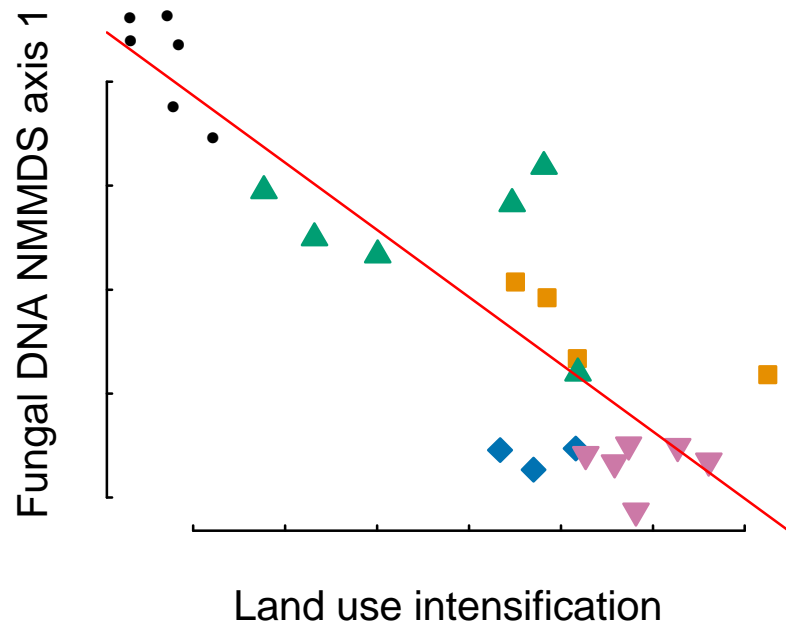
1. Define your question
2. Design sampling
3. Collect samples
4. Extract DNA
5. Sequence DNA
6. **Match DNA to species**



# How to assess biodiversity using eDNA?

## 7 fundamental steps

1. Define your question
2. Design sampling
3. Collect samples
4. Extract DNA
5. Sequence DNA
6. Match DNA to species
- 7. Analyse data to answer question**





# Biodiversity assessment using eDNA

## 7 fundamental steps

1. Define your question **HARD**
2. Design sampling **MODERATE**
3. Collect samples **EASY**
4. Extract DNA **EASY**
5. Sequence DNA **EASY**
6. Match DNA to species **HARD**
7. Analyse data to answer question **MODERATE**

# How to assess biodiversity using eDNA?

## 7 fundamental steps

- 1. Define your question**
2. Design sampling
3. Collect samples
4. Extract DNA
5. Sequence DNA
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# 1. Define your question

Sets the spatial scale of the study

- What is the fungal community in my paddock?
- How does land use affect fungal community composition?

Determines sample media and target taxonomic groups

- What fungal endophyte species occur on kiwifruit leaves?
- How does cultivation affect the community composition of soil biota?

# Main question from the Next-Generation Biodiversity Assessment Project (Phase 1)

How does the richness and composition of biological communities vary within and across land uses?

**Natural forest**



**Planted forest**



**Low-producing  
grassland**



**High-producing  
grassland**



**Vineyard**



# Biodiversity assessment using eDNA

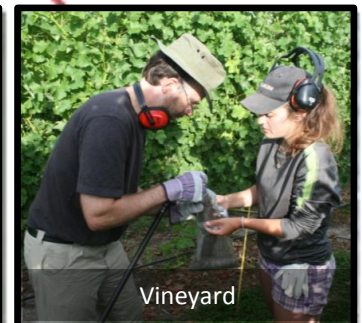
## 7 basic steps

1. Define your question ✓
2. **Design sampling**
3. Collect samples
4. Extract DNA
5. Sequence DNA
6. Match DNA to species
7. Analyse data to answer question



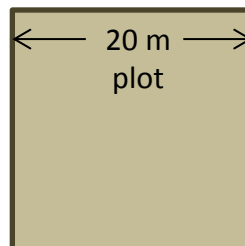
# Phase 1: Proof-of-concept in a model catchment (Wairau Valley, Marlborough)

5 land use types along an intensification gradient



- Based on national grid of 20x20m plots used by DoC, MfE and regional councils
- Traditional plot-based measurements (plants, birds) alongside molecular methods

Field-based  
Bird and plant surveys



Soil cores for molecular analysis



# Biodiversity assessment using eDNA

## 7 fundamental steps

1. Define your question ✓
2. Design sampling ✓
- 3. Collect samples**
4. Extract DNA
5. Sequence DNA
6. Match DNA to species
7. Analyse data to answer question

## 3. Collect samples

- Landowner and industry permissions
- Iwi consultation
- Logistical considerations

However, eDNA sample collection is relatively straightforward and could be done by the farmers themselves



# Biodiversity assessment using eDNA

## 7 fundamental steps

1. Define your question ✓
2. Design sampling ✓
3. Collect samples ✓
4. **Extract DNA**
5. Sequence DNA
6. Match DNA to species
7. Analyse data to answer question



## 4. Extract DNA

- Commercially available kits
- Two distinct stages
  - Isolate DNA from sample
  - Use PCR to amplify target gene regions
- PCR primer selection is critical



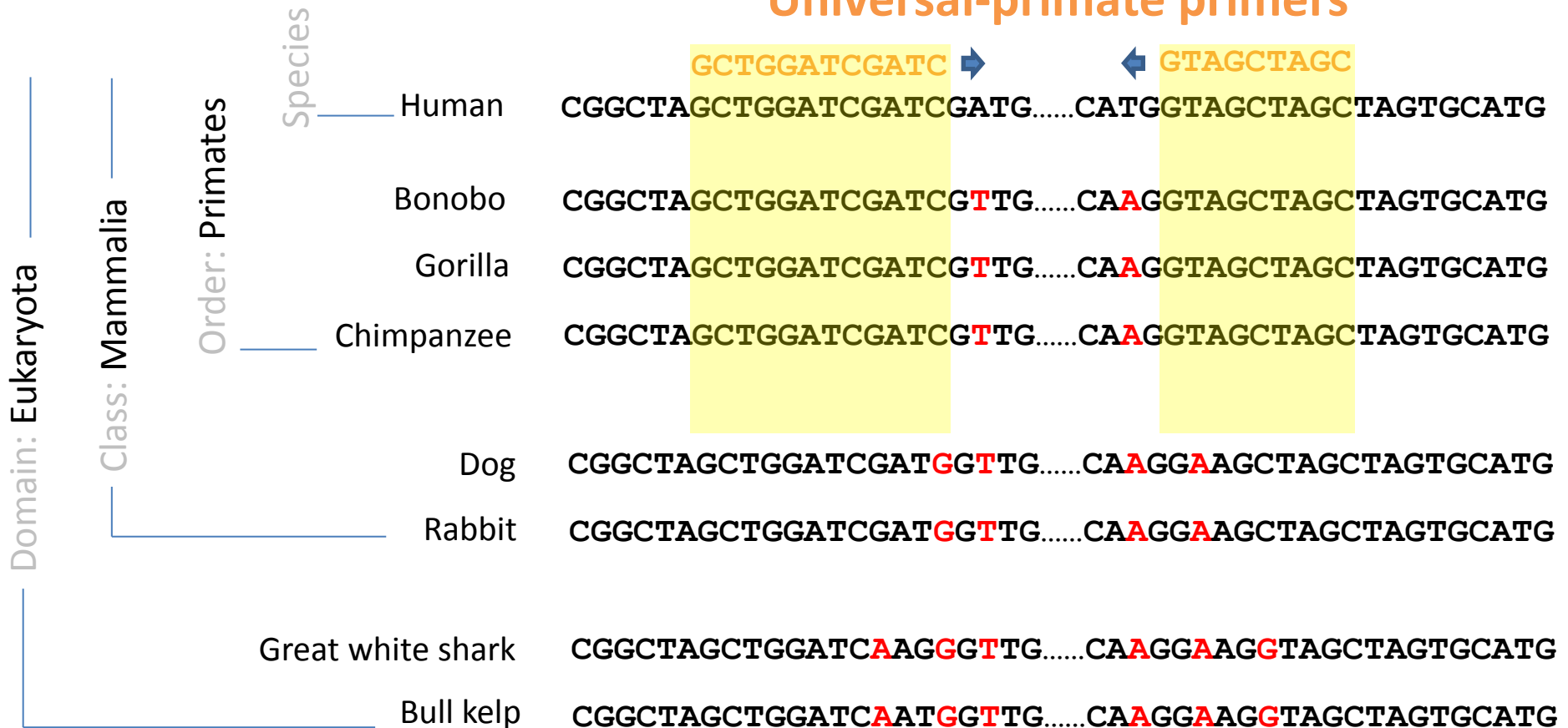
# General vs specific primers

## Human-specific primers

Domain: Eukaryota	Class: Mammalia	Order: Primates	Species	Human	CGGCTAGCT <b>TGGATCGATCGA</b> ..... <b>TGGTAGCTA</b> CATGGTAGCTAGCTAGTGCATG
			Bonobo	CGGCTAGCTGGATCGATCG <b>TTG</b> .....CA <b>AGG</b> TAGCTAGCTAGTGCATG	
			Gorilla	CGGCTAGCTGGATCGATCG <b>TTG</b> .....CA <b>AGG</b> TAGCTAGCTAGTGCATG	
			Chimpanzee	CGGCTAGCTGGATCGATCG <b>TTG</b> .....CA <b>AGG</b> TAGCTAGCTAGTGCATG	
		Dog	CGGCTAGCTGGATCGAT <b>GGTTG</b> .....CA <b>AGGAAG</b> CTAGCTAGTGCATG		
		Rabbit	CGGCTAGCTGGATCGAT <b>GGTTG</b> .....CA <b>AGGAAG</b> CTAGCTAGTGCATG		
		Great white shark	CGGCTAGCTGGATC <b>AAGGGTTG</b> .....CA <b>AGGAAGG</b> TAGCTAGTGCATG		
		Bull kelp	CGGCTAGCTGGATC <b>AATGGTTG</b> .....CA <b>AGGAAGG</b> TAGCTAGTGCATG		

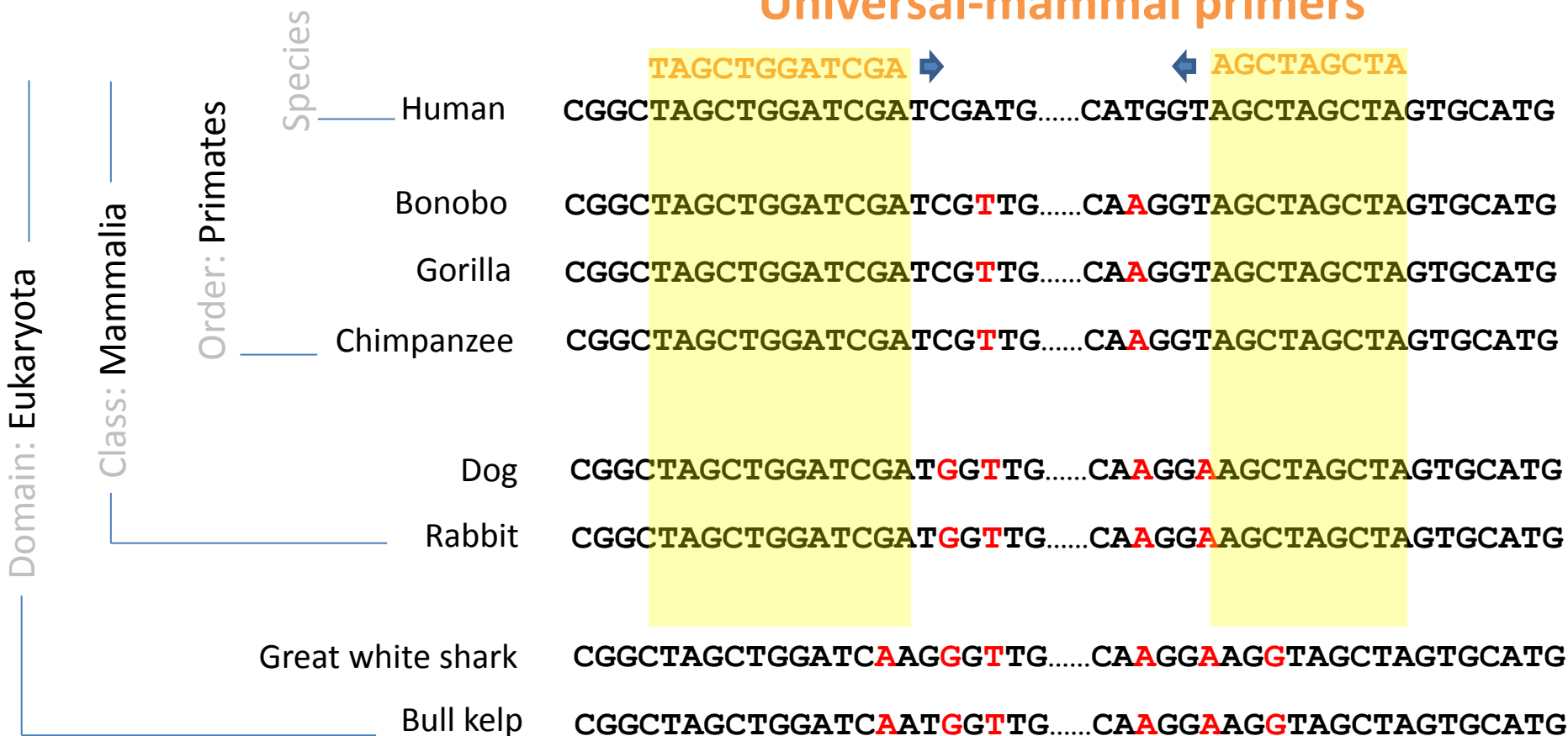
# General vs specific primers

## Universal-primate primers



# General vs specific primers

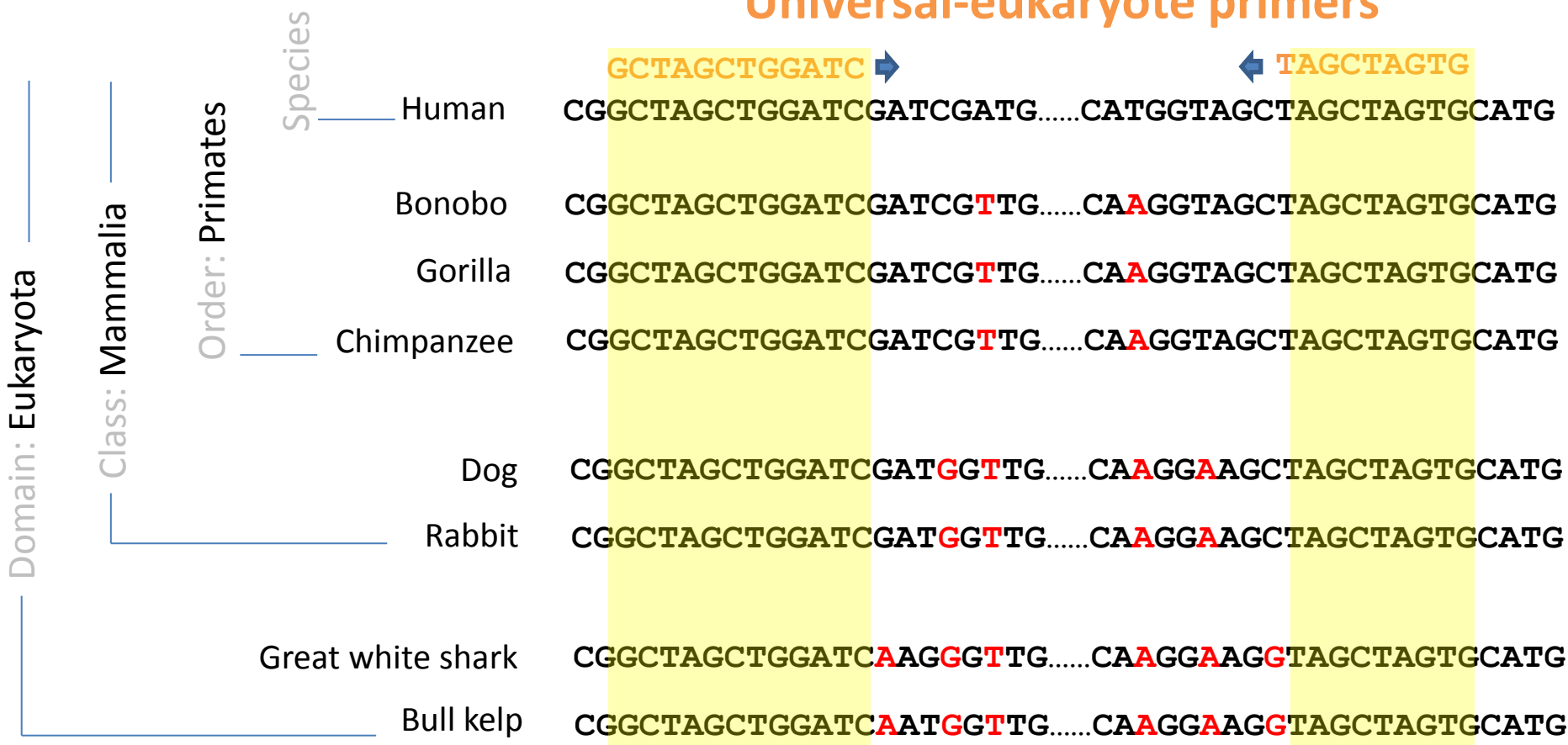
## Universal-mammal primers





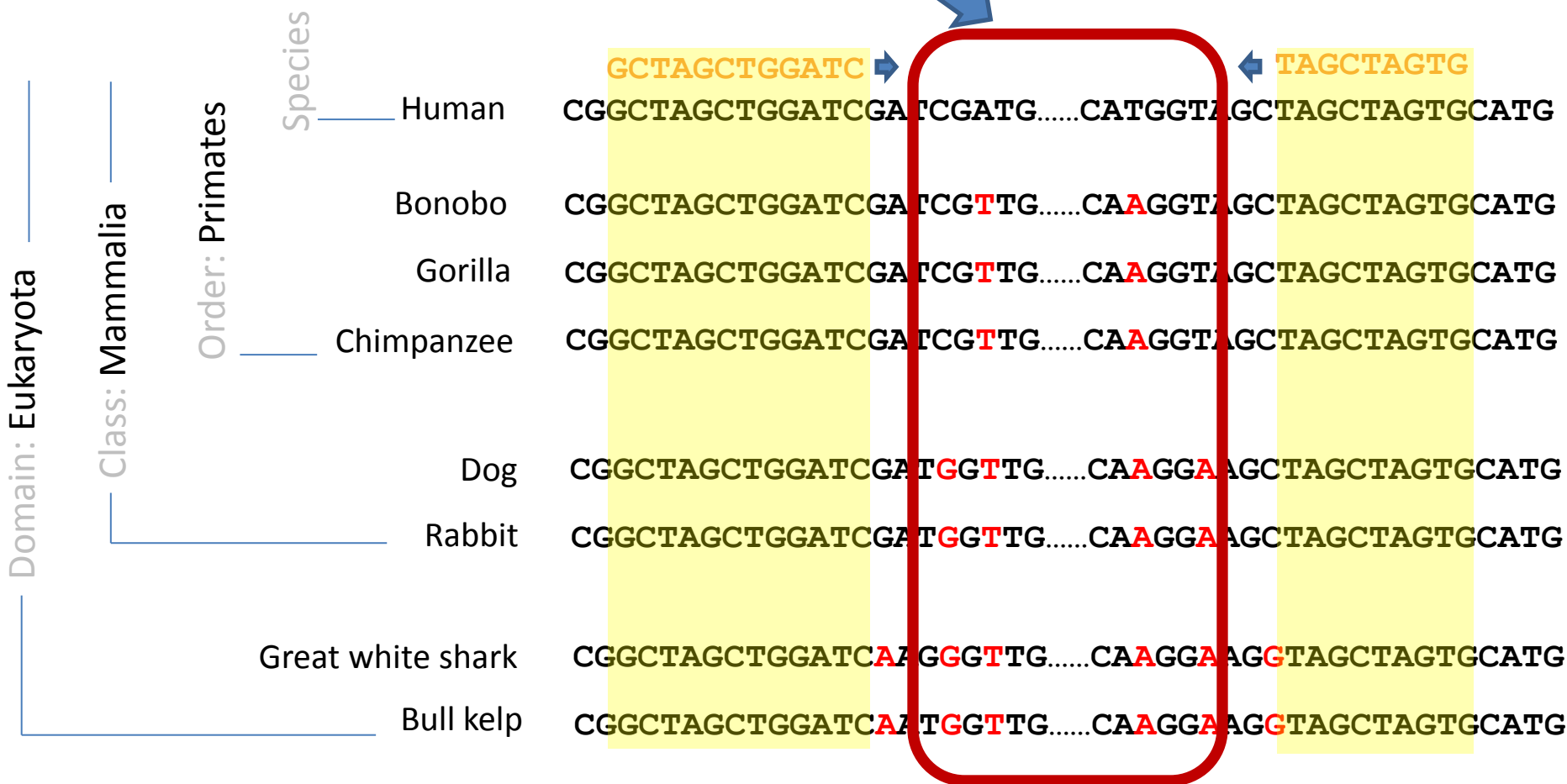
# General vs specific primers

## Universal-eukaryote primers



# Ability of a primer to differentiate species

## Variability in amplified gene region



# Biodiversity assessment using eDNA

## 7 fundamental steps

1. Define your question ✓
2. Design sampling ✓
3. Collect samples ✓
4. Extract DNA ✓
5. **Sequence DNA**
6. Match DNA to species
7. Analyse data to answer question

## 5. Sequence DNA

- Next-generation sequencing platforms
- Commercially available service
- Can be done on a pooled DNA sample
- Technology rapidly evolving
- Big data outputs





# Biodiversity assessment using eDNA

## 7 fundamental steps

1. Define your question ✓
2. Design sampling ✓
3. Collect samples ✓
4. Extract DNA ✓
5. Sequence DNA ✓
6. **Match DNA to species**
7. Analyse data to answer question

## 6. Match DNA to species (or OTUs)

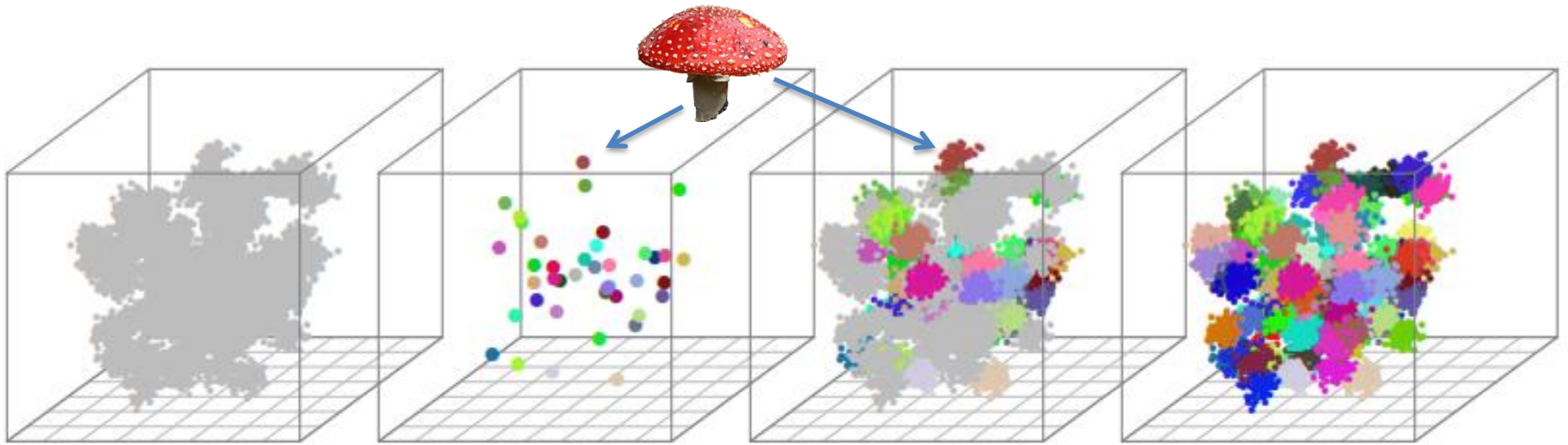
Operational taxonomic units (OTUs) allow identification of 'species' without taxonomic matches

Assigning species names to OTU's is reliant on reference databases

– Genbank, GreenGenes

Going from DNA to OTU's &/or species is **challenging**

- Incomplete reference data
- Which bioinformatics method to use?
- What taxonomic resolution is most appropriate?

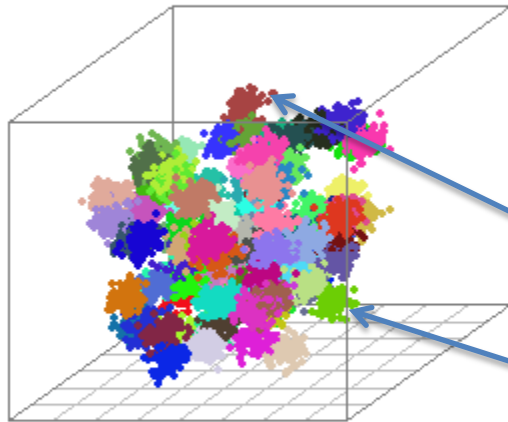


**1:** start with DNA data in N-dimensional similarity cloud

**2:** “Seed” space with known species from reference library (e.g., from a specimen)

**3:** Match unknown data to these known references

**4:** Form new “unknown” clusters where no known species match



**5:** Assign names & identity based on:  
 - known “seed” that formed cluster  
 OR  
 - nearest known species in database (to some level of confidence)

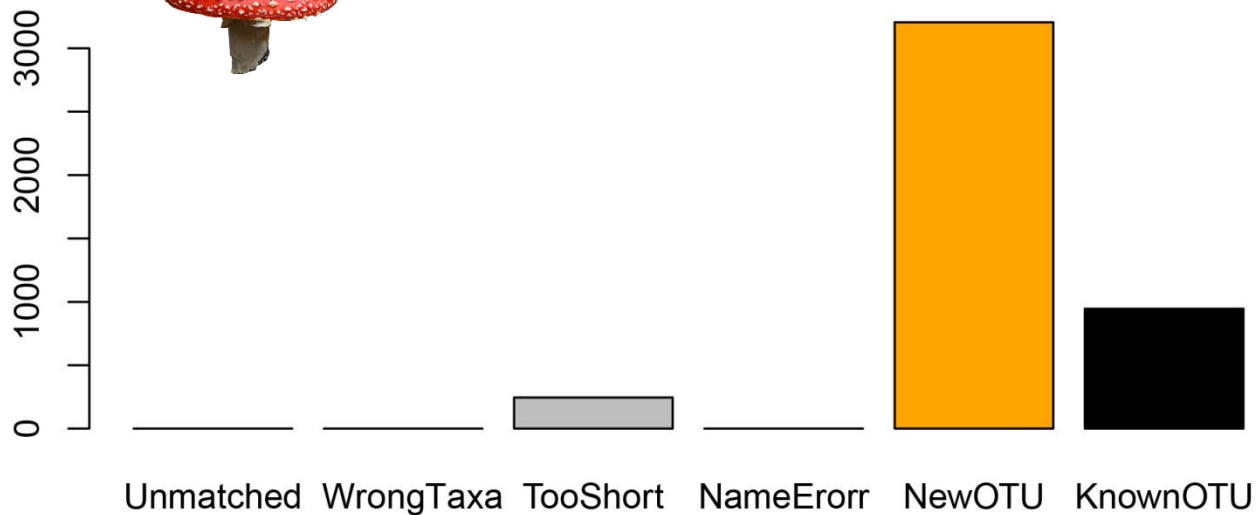
*Amanita muscaria* (OTU matched to known species)

Unknown rust fungus (OTU family level match)

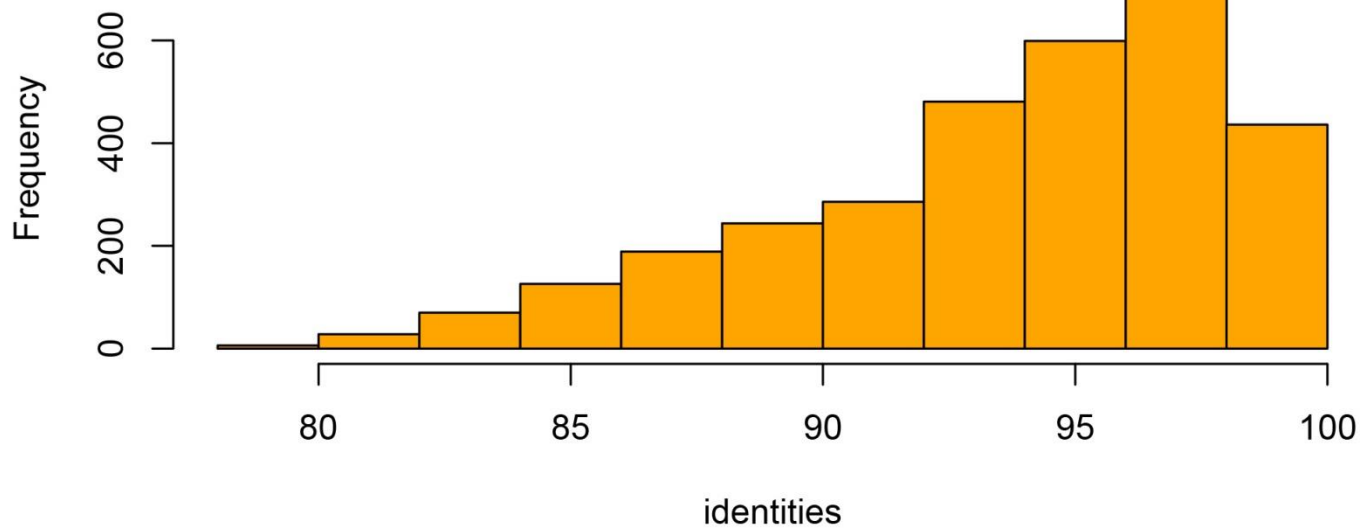




## Fungi ITS1F OTU matching

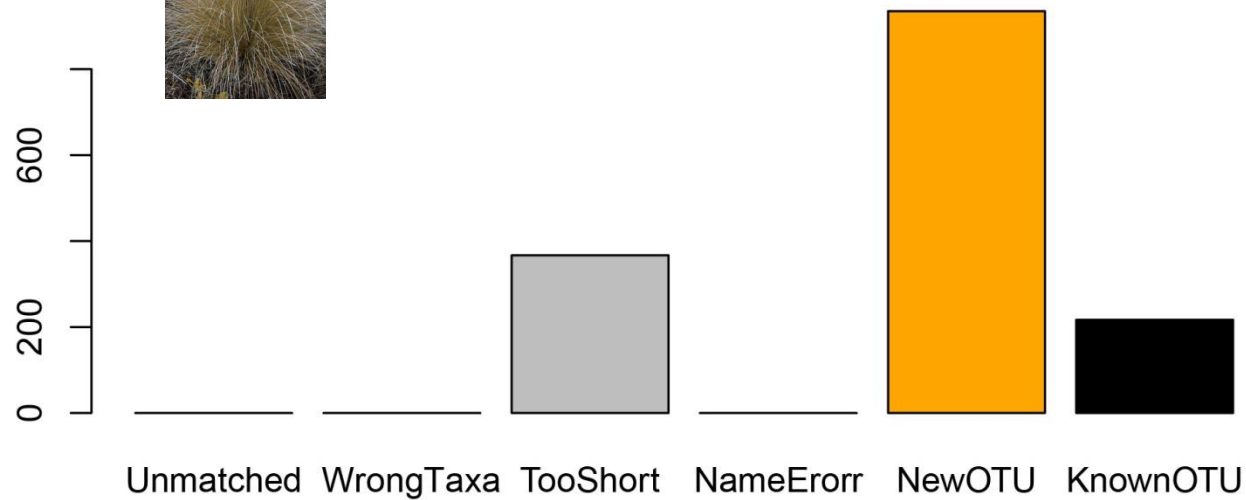


## New OTUs %match to Known

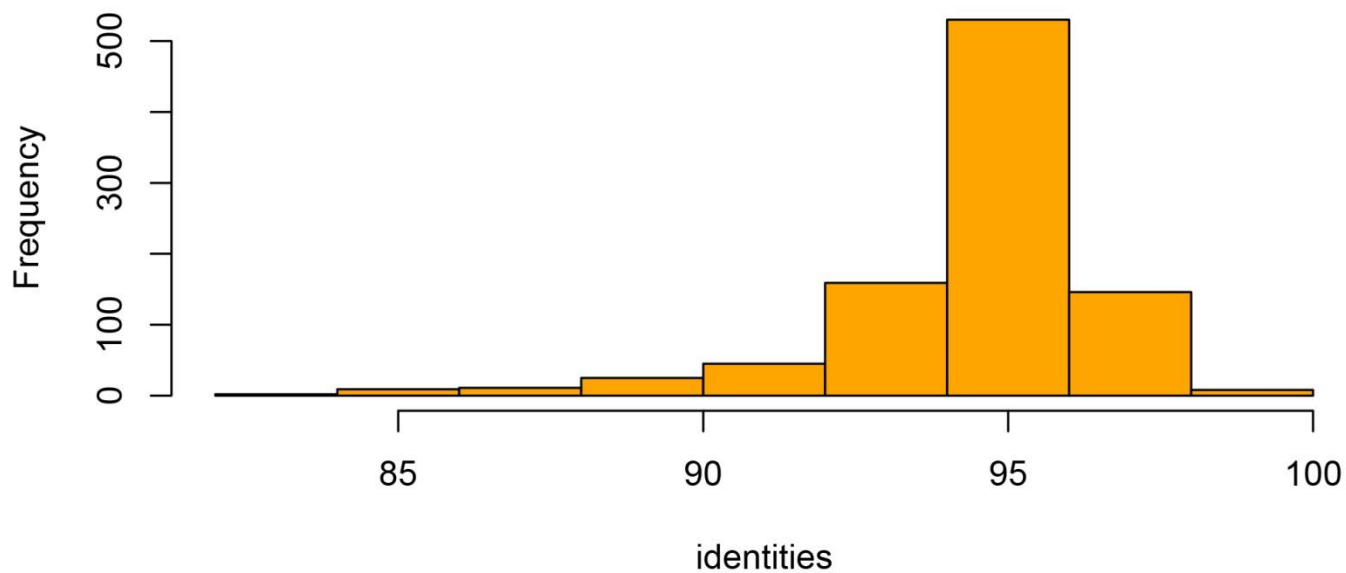




## Plants OTU matching



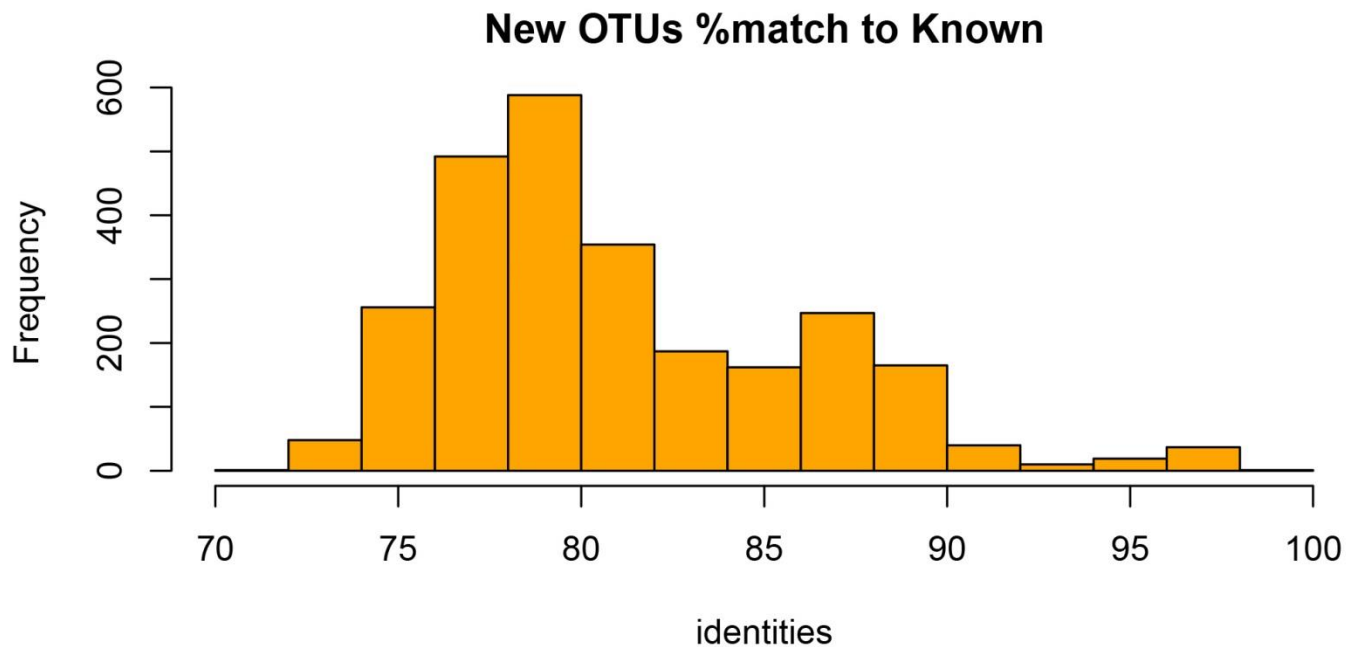
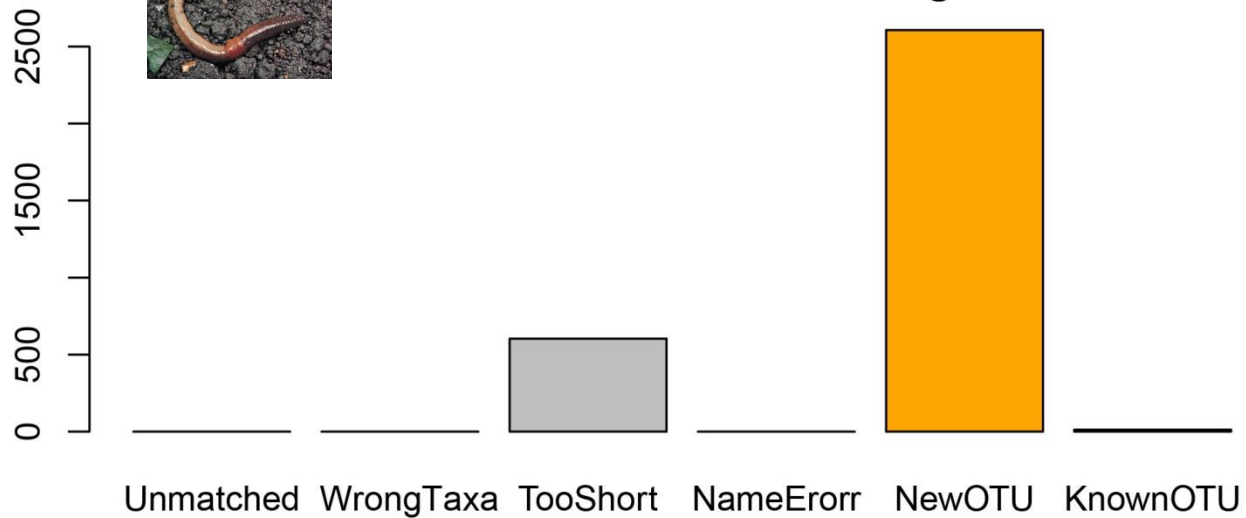
## New OTUs %match to Known







## Invertebrates OTU matching

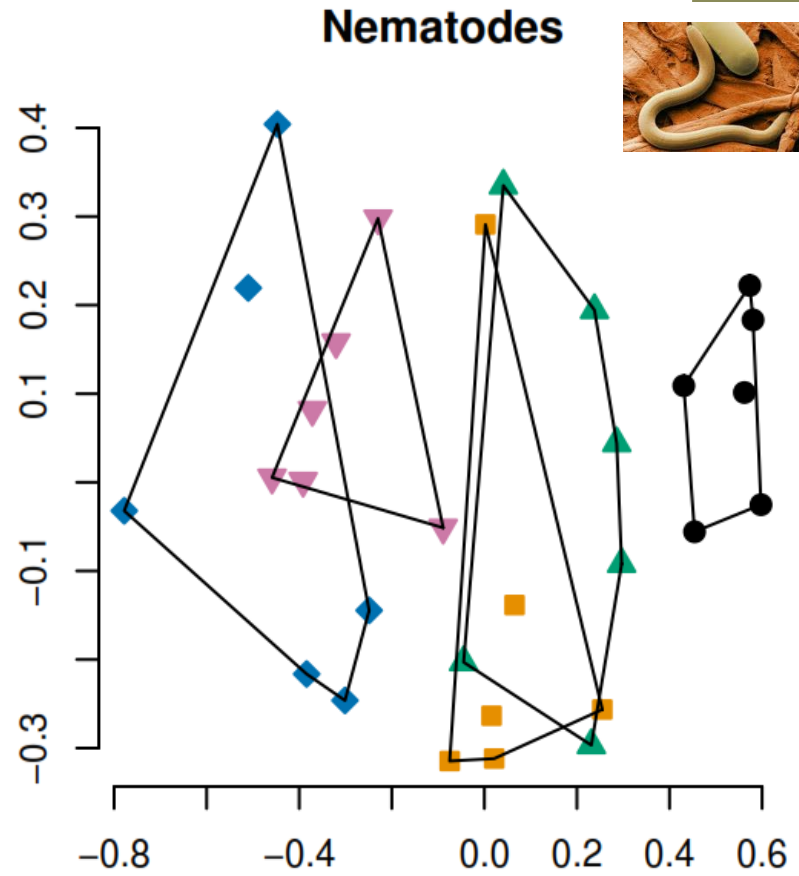
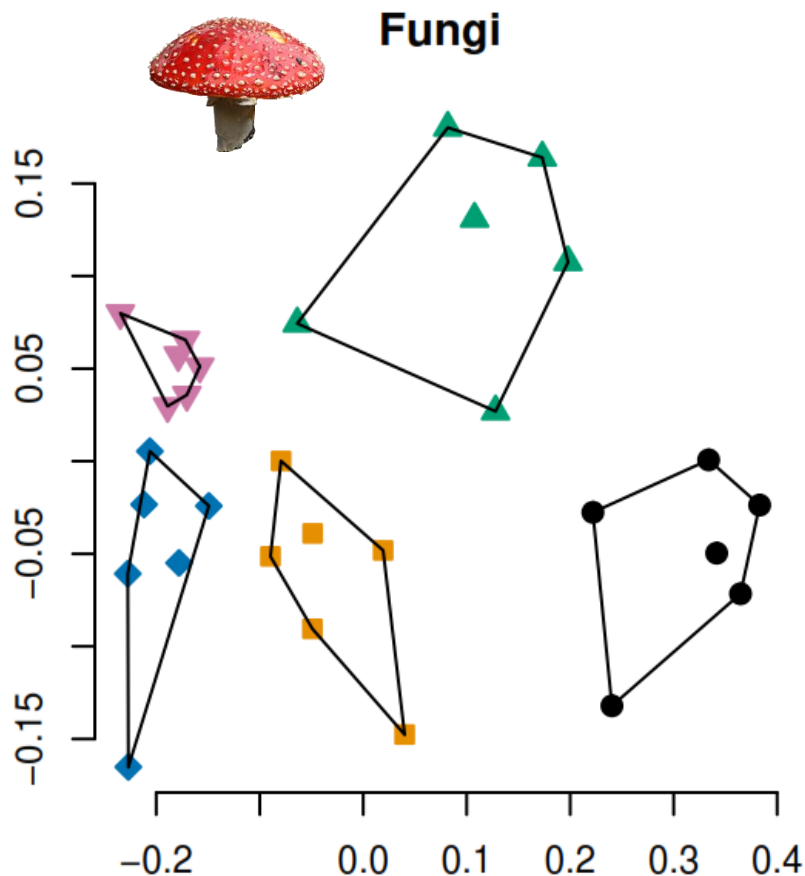


# Biodiversity assessment using eDNA

## 7 fundamental steps

1. Define your question ✓
2. Design sampling ✓
3. Collect samples ✓
4. Extract DNA ✓
5. Sequence DNA ✓
6. Match DNA to species ✓
- 7. Analyse data to answer question**

# How does the **composition** of biological communities vary with land use?

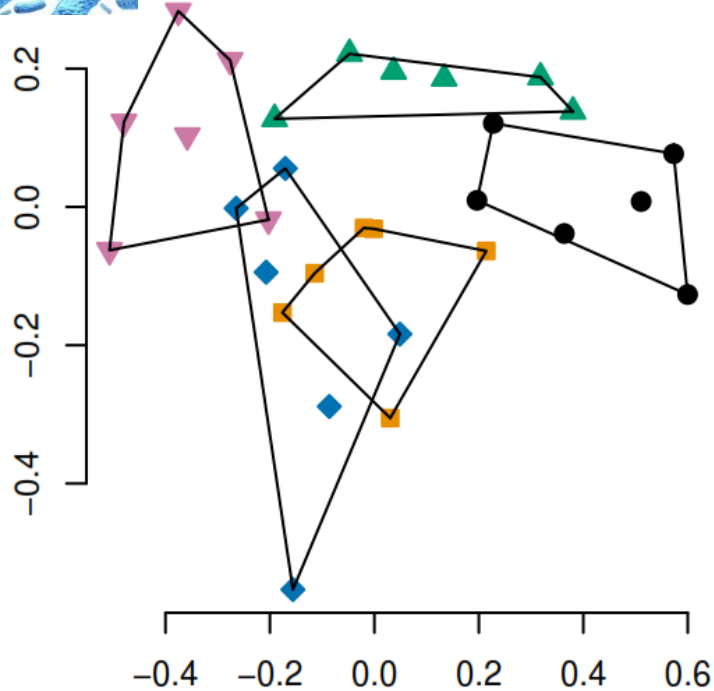


- Native forest
- ▲ Pine forest
- Low producing grassland
- ◆ High producing grassland
- ▼ Vineyard

# How does the **composition** of biological communities vary with land use?

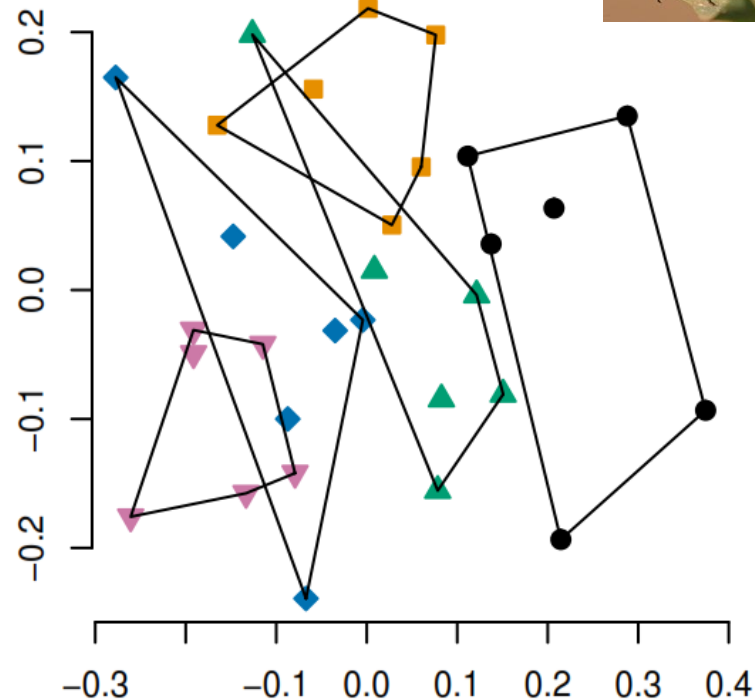


### Bacteria



- Native forest
- ▲ Pine forest

### Arthropods



- Low producing grassland
- ◆ High producing grassland
- ▼ Vineyard

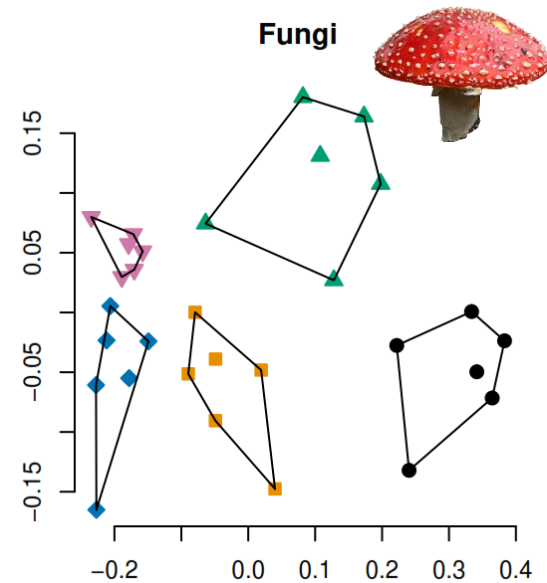
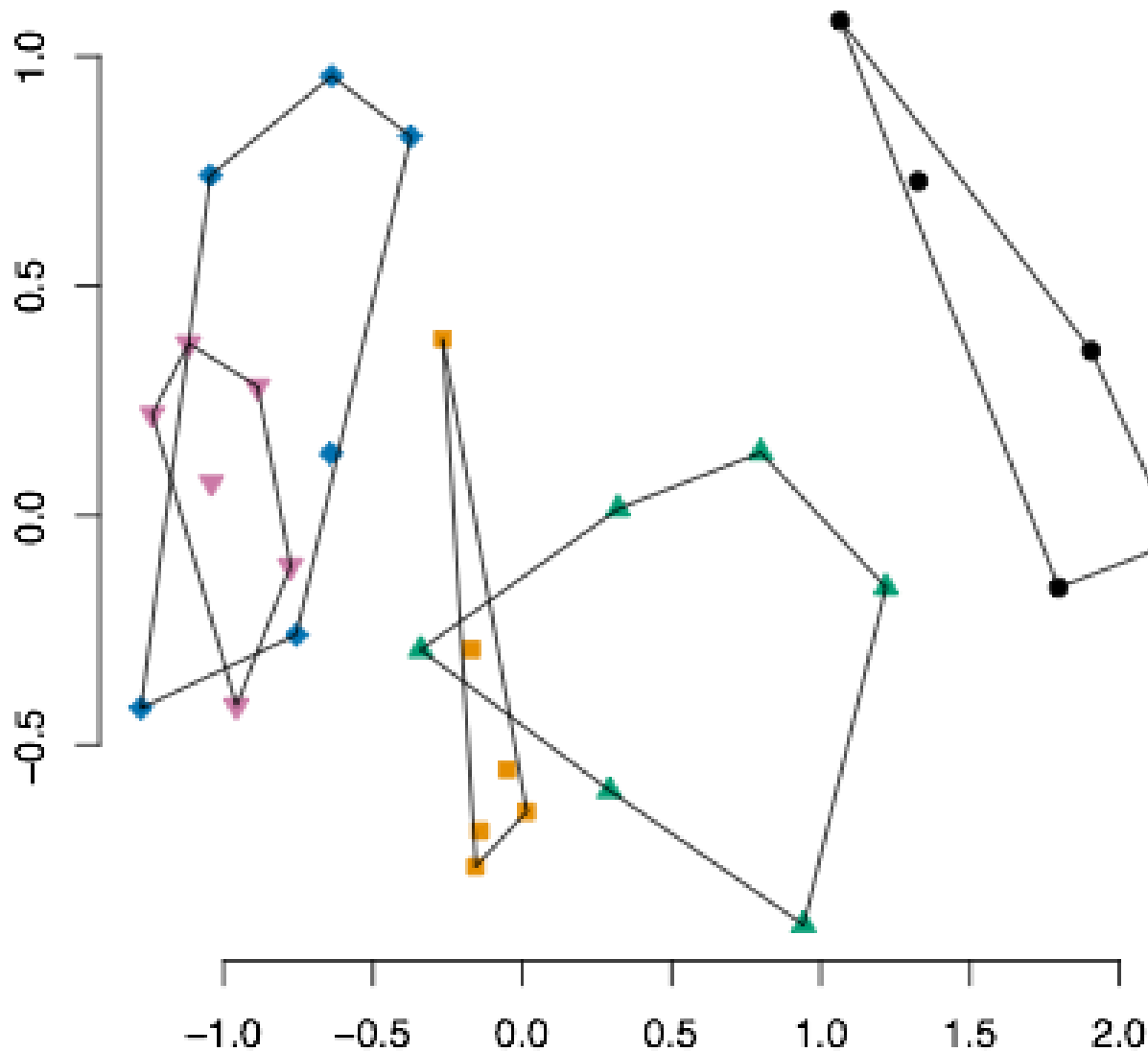


# Plant community

(based on non-DNA field measurements)

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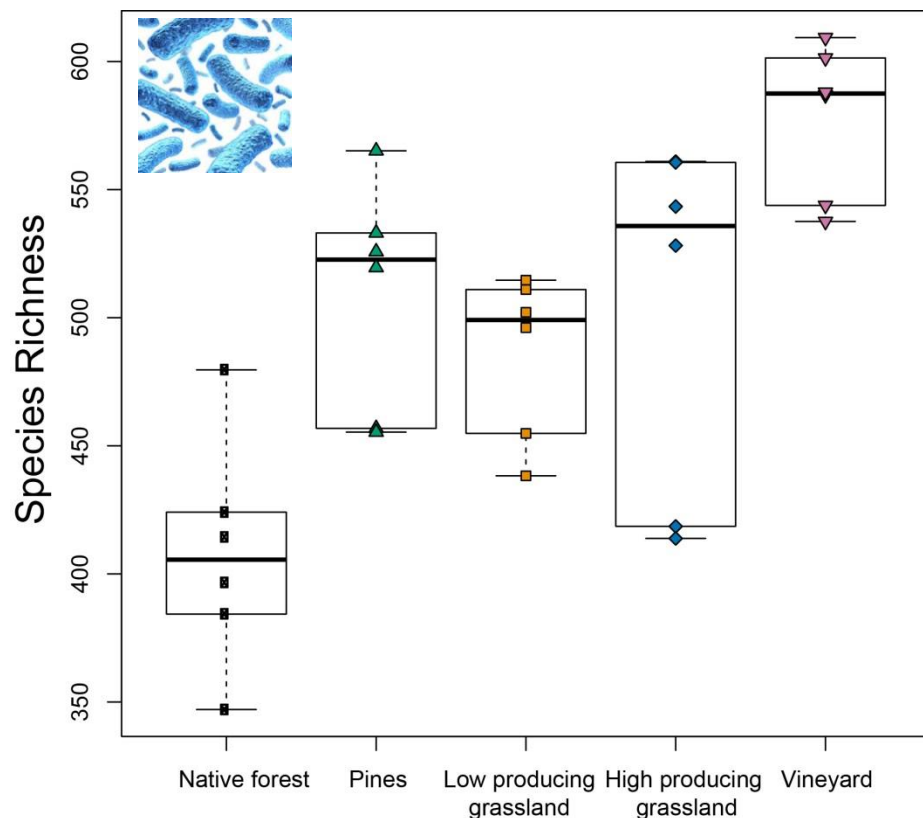


- Low producing grassland
- High producing grassland
- Vineyard
- Native forest
- Pine forest

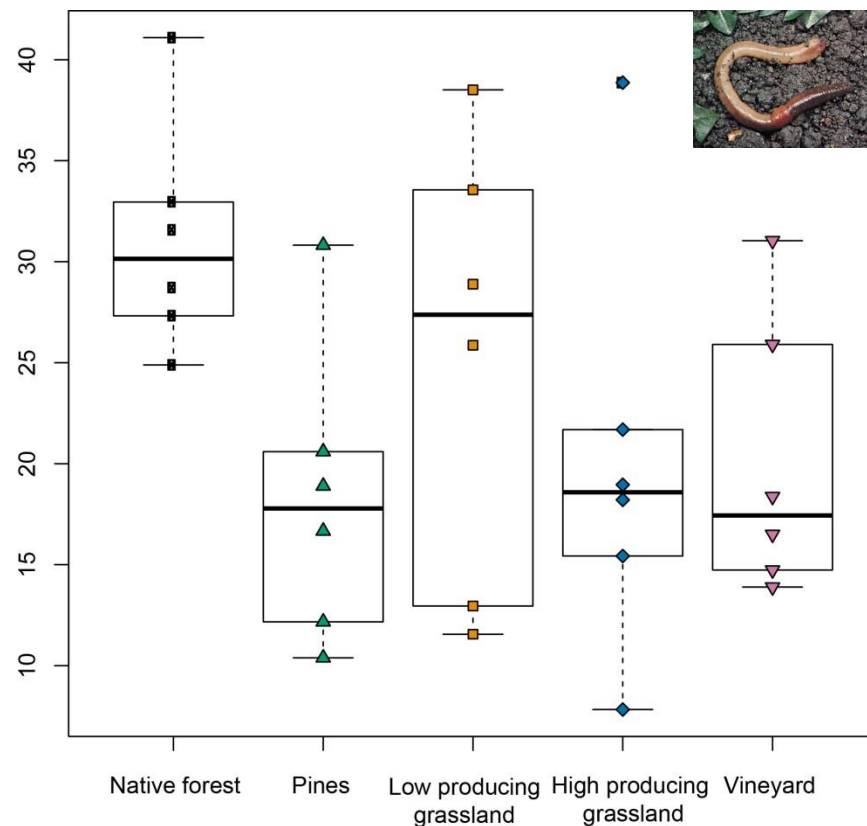


# How does the **richness** of biological communities vary with land use?

## Bacteria



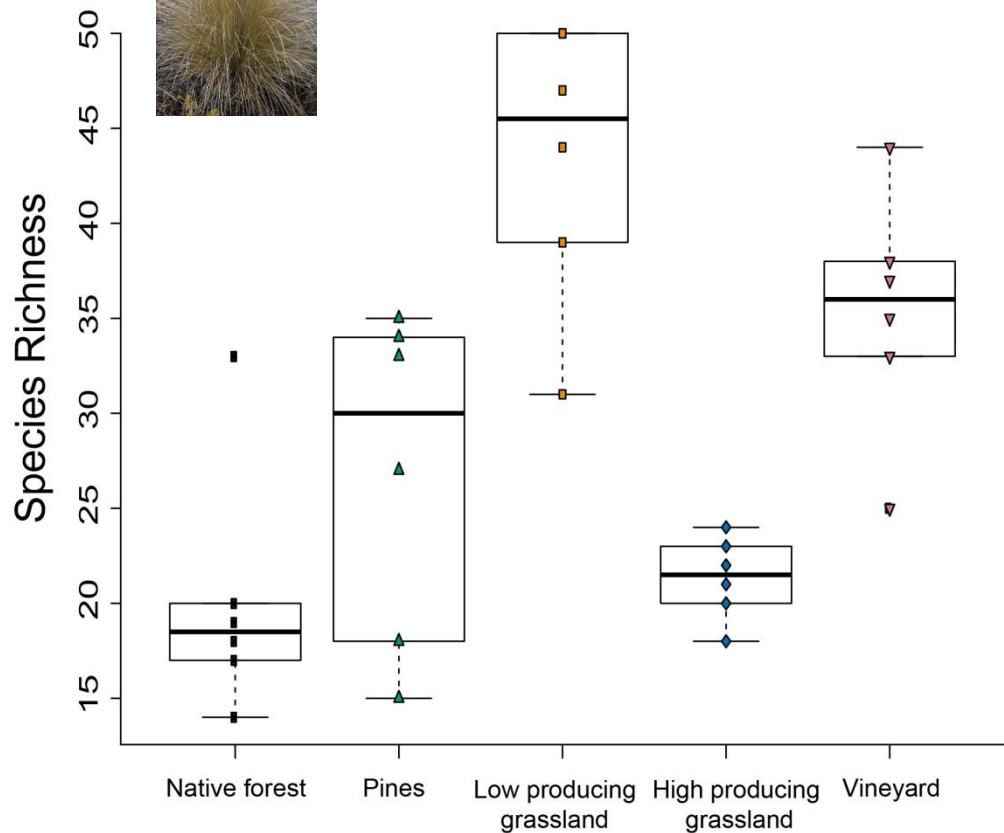
## Invertebrates



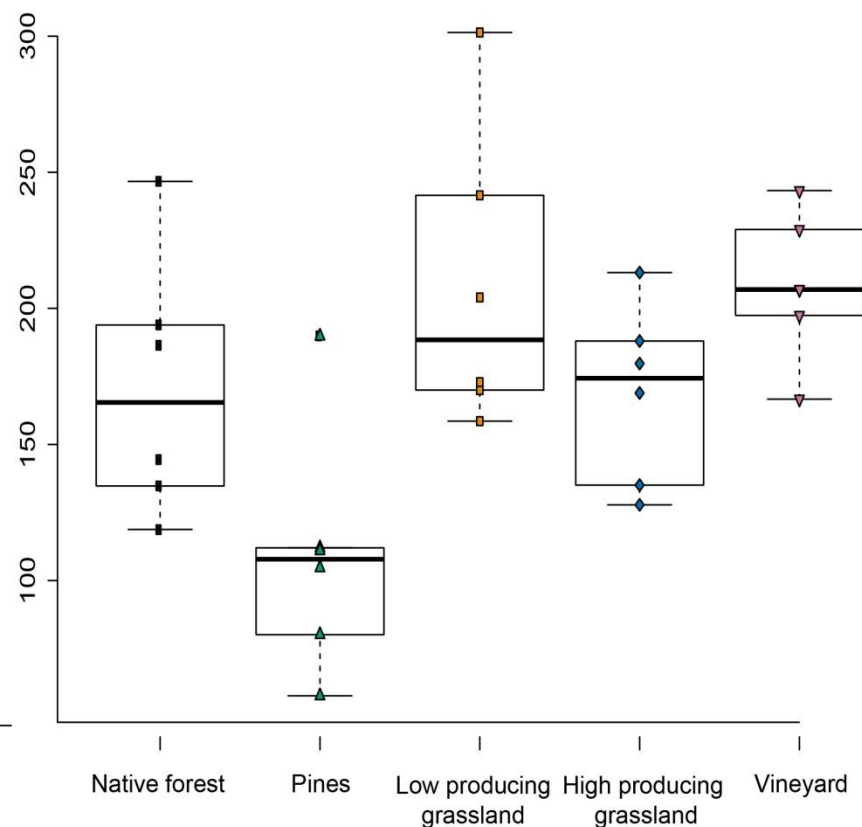
Land use type

# How does the **richness** of biological communities vary with land use?

## Plants

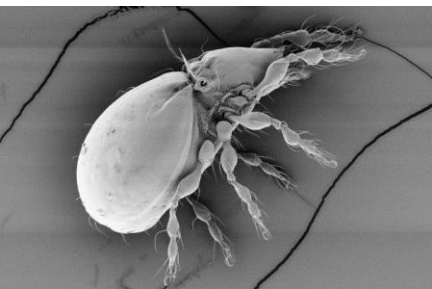


## Fungi



# Primary sector eDNA applications

- Demonstrating environmental performance
- Improving farm management
- Biosecurity detection and monitoring



# Application 1

## Demonstrating environmental performance

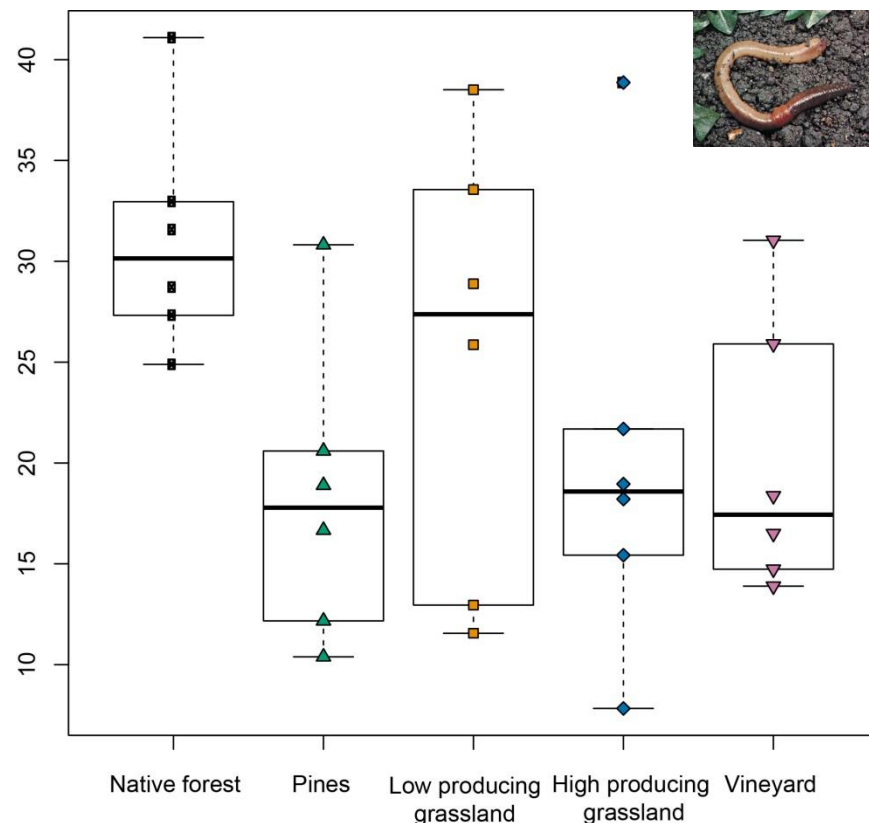
- Demonstrating positive effects of industry on biodiversity is an increasing market requirement
- Benchmarking within sector groups provides a framework to improve management practices
- eDNA derived indicators could be used to demonstrate maintenance (or enhancement) of whole-ecosystem biodiversity

# Application 1

## Demonstrating environmental performance

- (e.g.) Maintenance of invertebrate species richness
- Large variability within land use types
  - Environmental or management driven?

Invertebrate richness

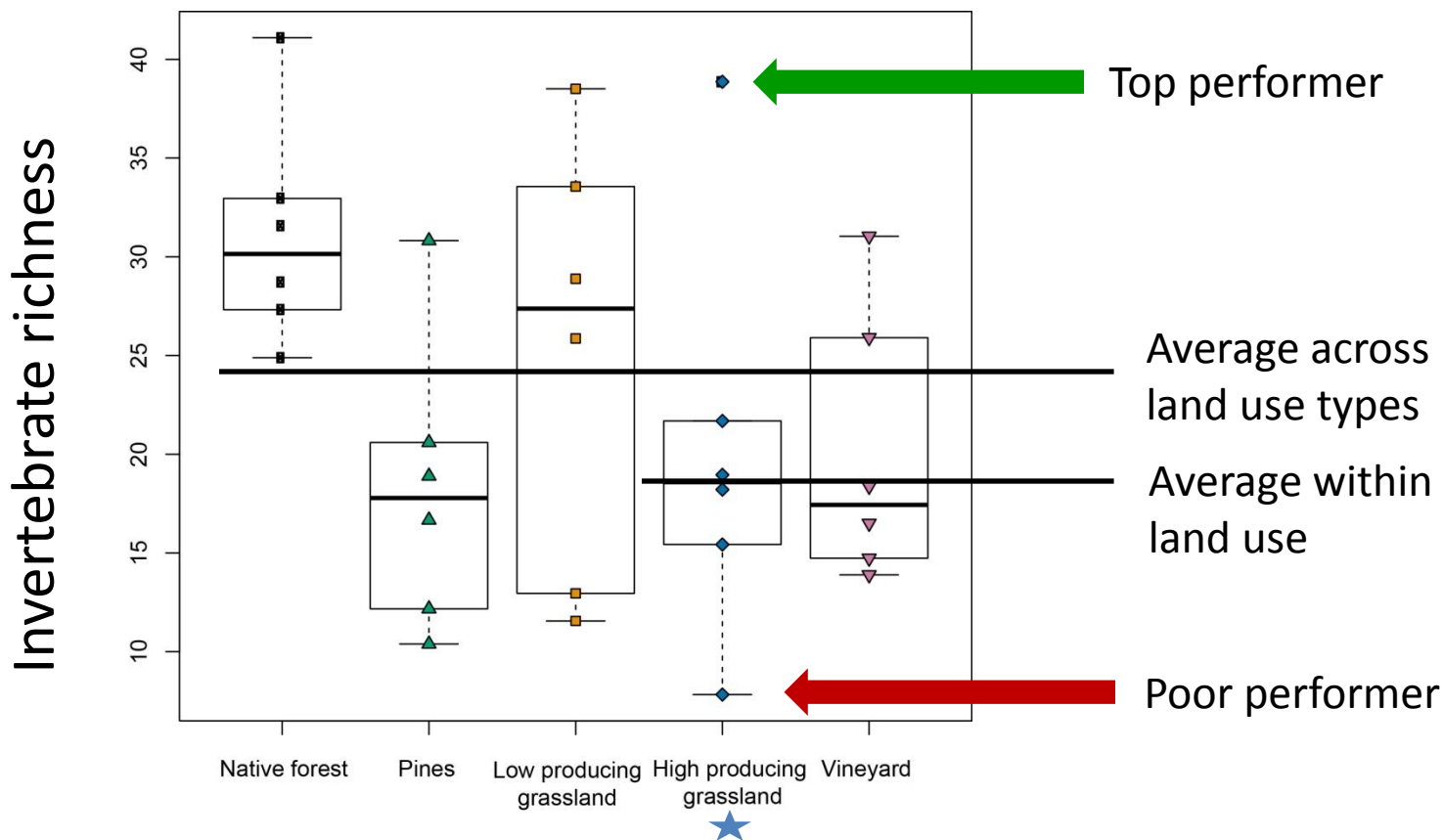




# Application 1

## Demonstrating environmental performance

- Benchmarking within and across land use



## Application 2

# Improving farm management

Future farming systems depend on healthy living soil

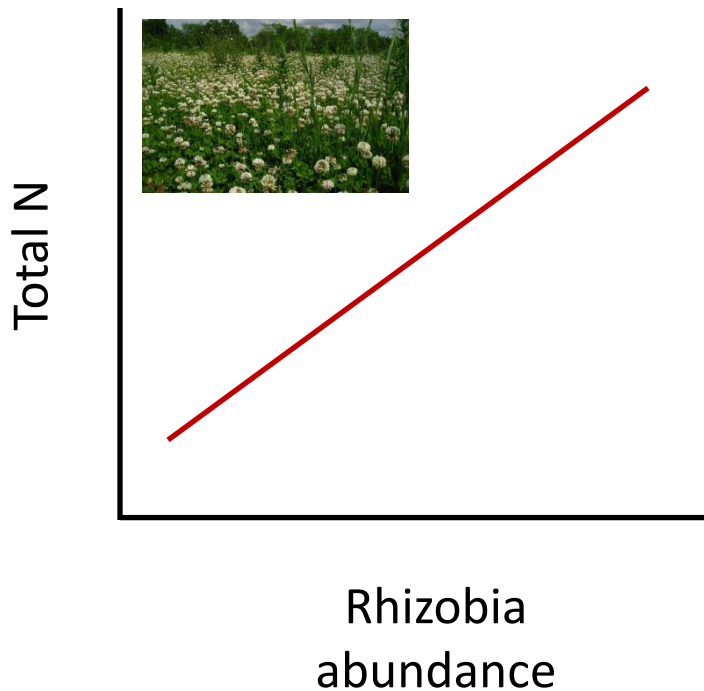
- Some biota in particular are known to be essential for productive systems
  - nitrogen fixing bacteria, mycorrhizal fungi, predatory mites
- eDNA can be used as a tool to understand the effect of soil biota (or the soil community) on farm productivity

# Application 2

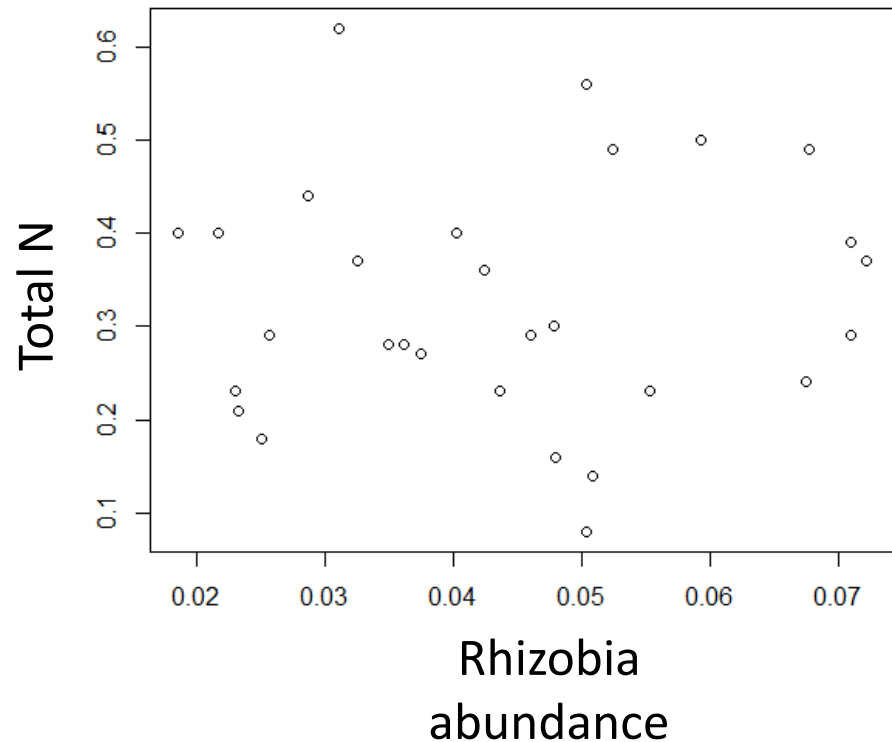
## Improving farm management

### Nitrogen fixing Rhizobia bacteria

Theory



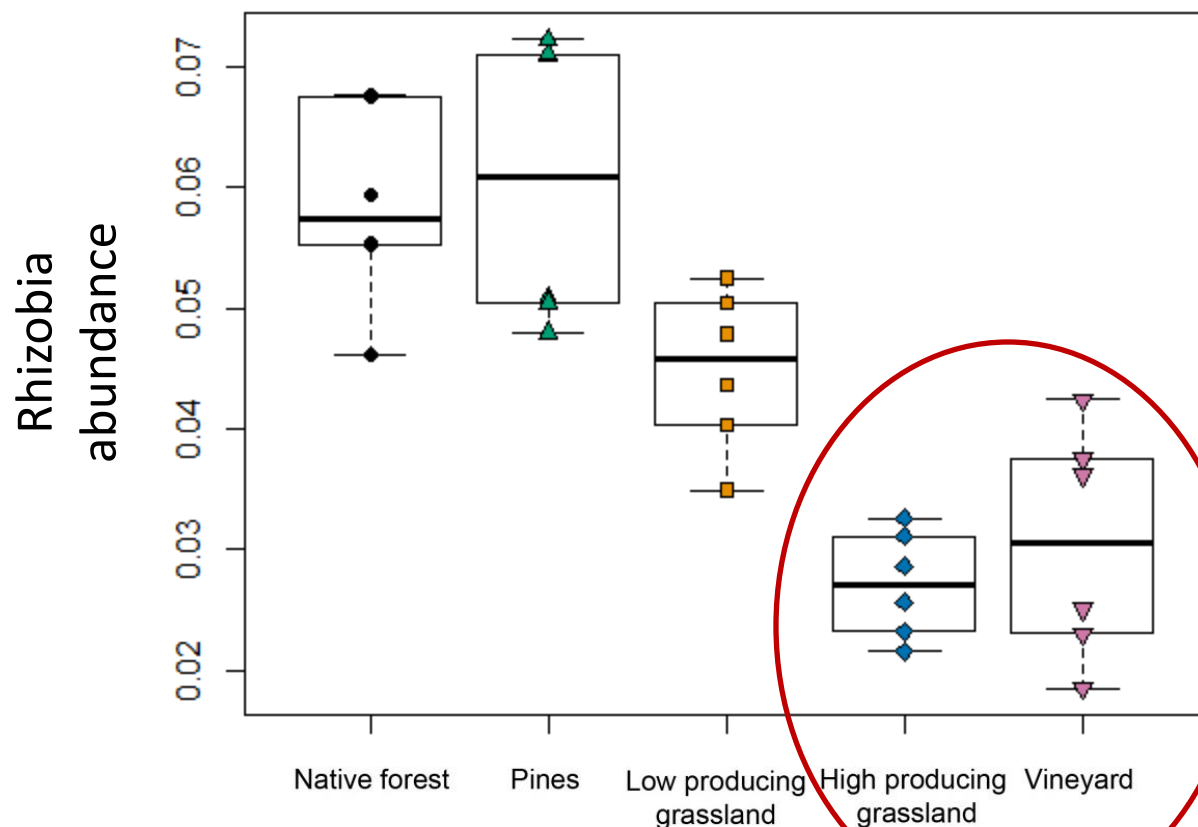
Phase 1 data



# Application 2

## Improving farm management

### Nitrogen fixing Rhizobia bacteria



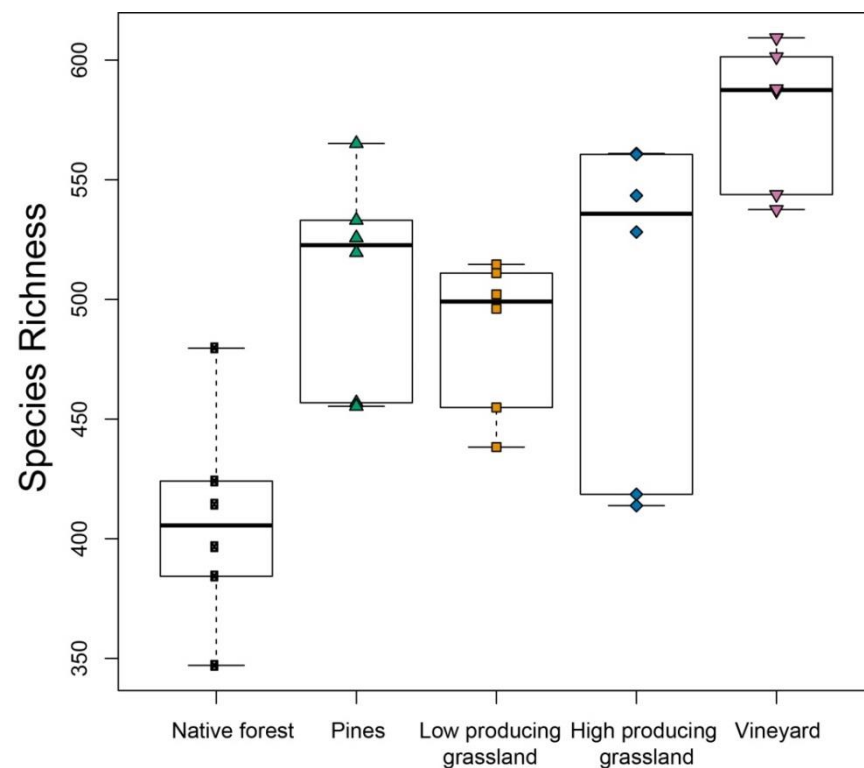
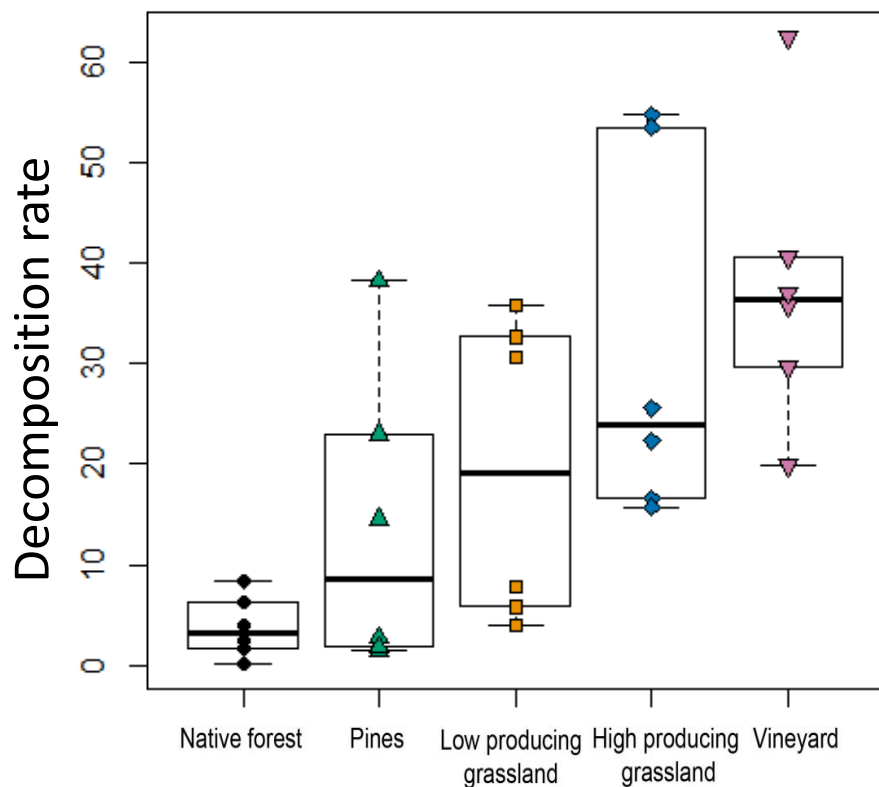
High N  
inputs  
from  
fertiliser

# Application 2

## Improving farm management

### Biotic controls of decomposition

#### Phase 1 (bacteria)



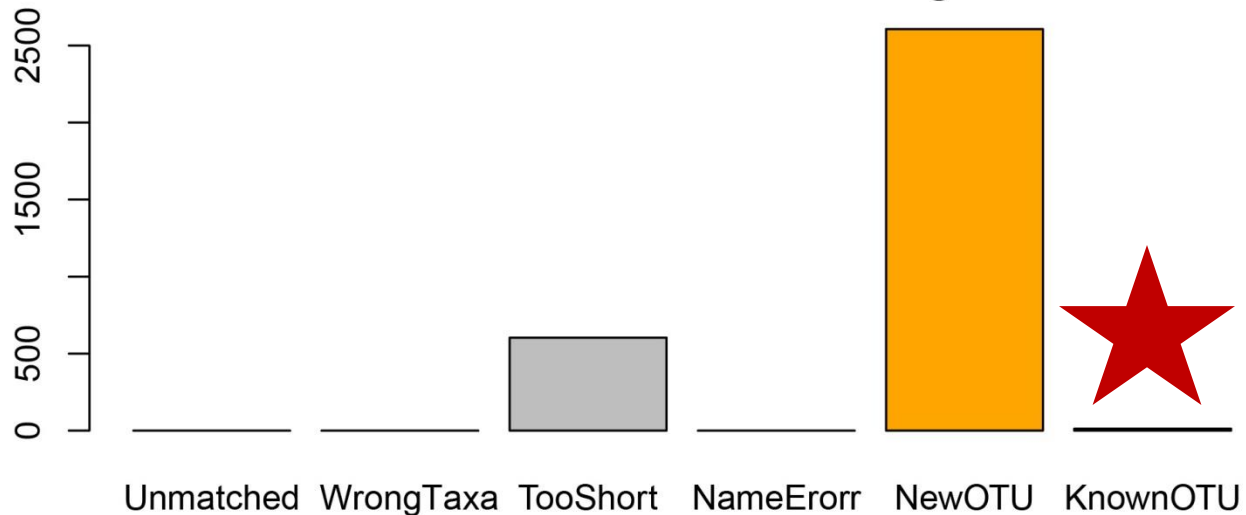


## Application 3

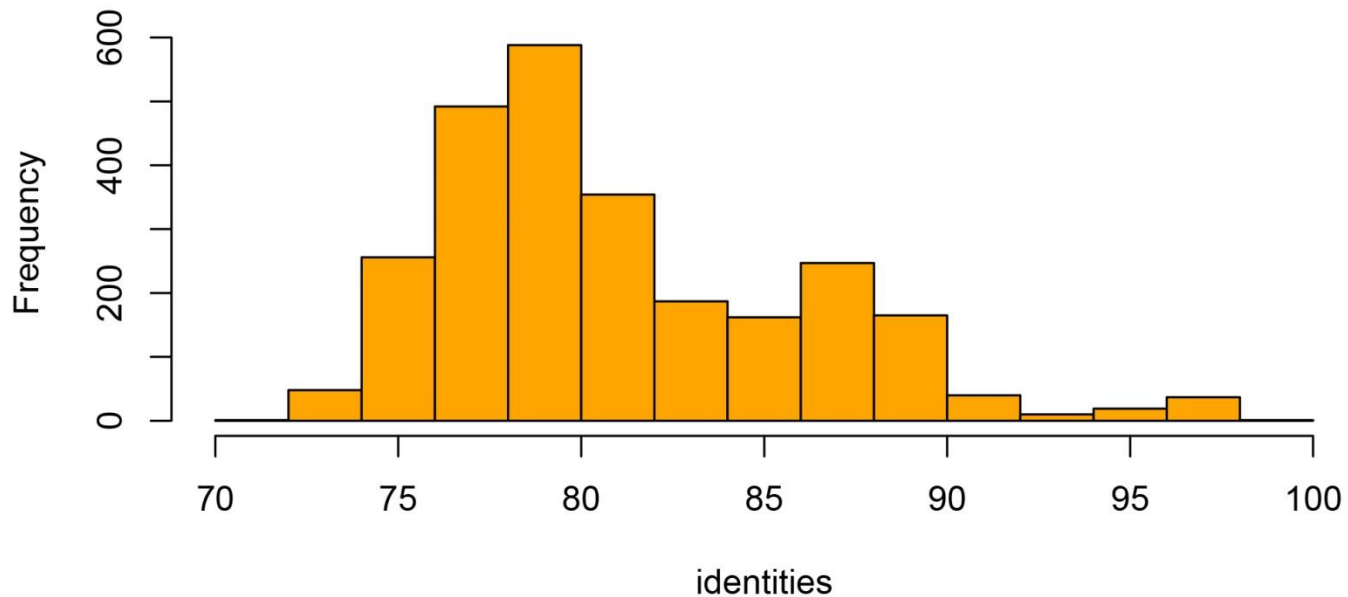
# Biosecurity detection and monitoring

- Detecting presence of a threat before the symptoms appear
  - e.g. detecting kauri dieback before trees die
- Requires better reference databases for many taxa

### Invertebrates OTU matching



### New OTUs %match to Known

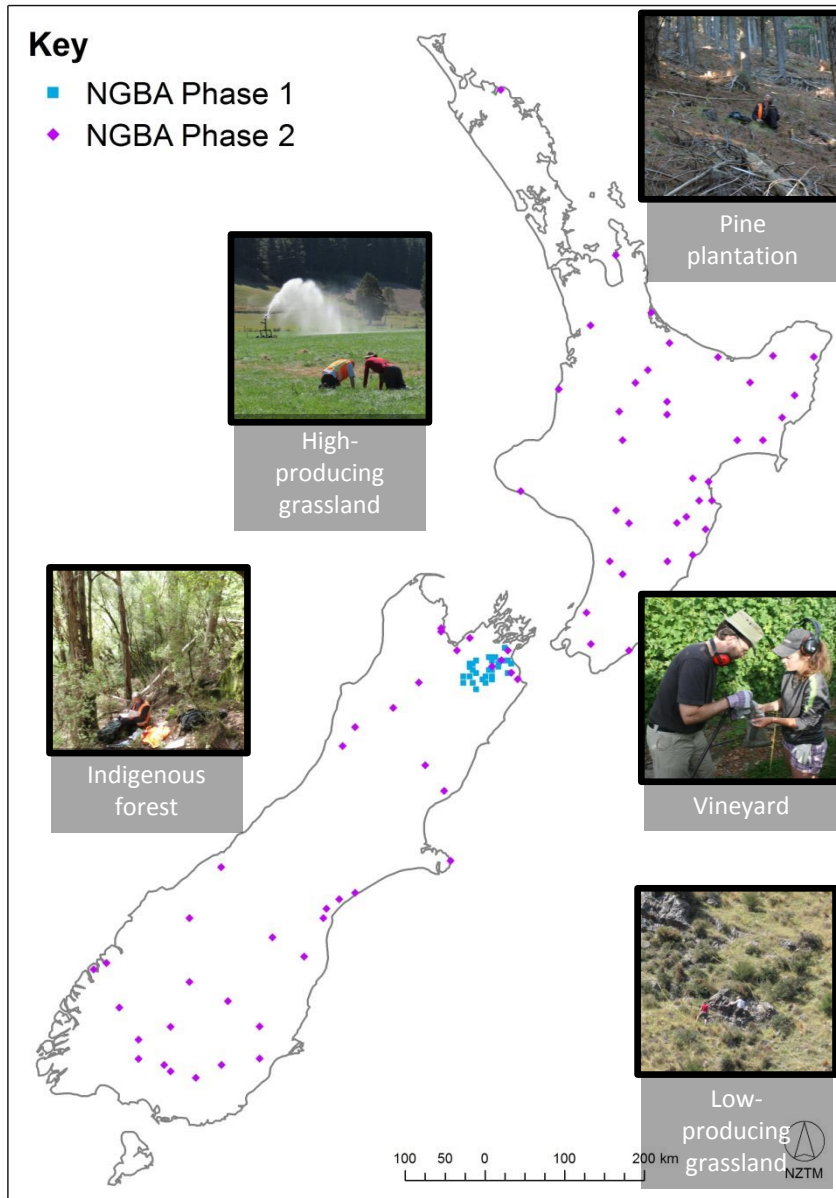


# Future directions

- Phase 2 of MBIE Smart Idea
- Biological Heritage National Science Challenge eDNA project



# Phase 2 – National & local variability







































14/01/2015









# National **SCIENCE** Challenges

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## MISSION

Reverse the decline of New Zealand's biological heritage, through a national partnership to deliver step change in research innovation, globally-leading technologies, and community and sector action.

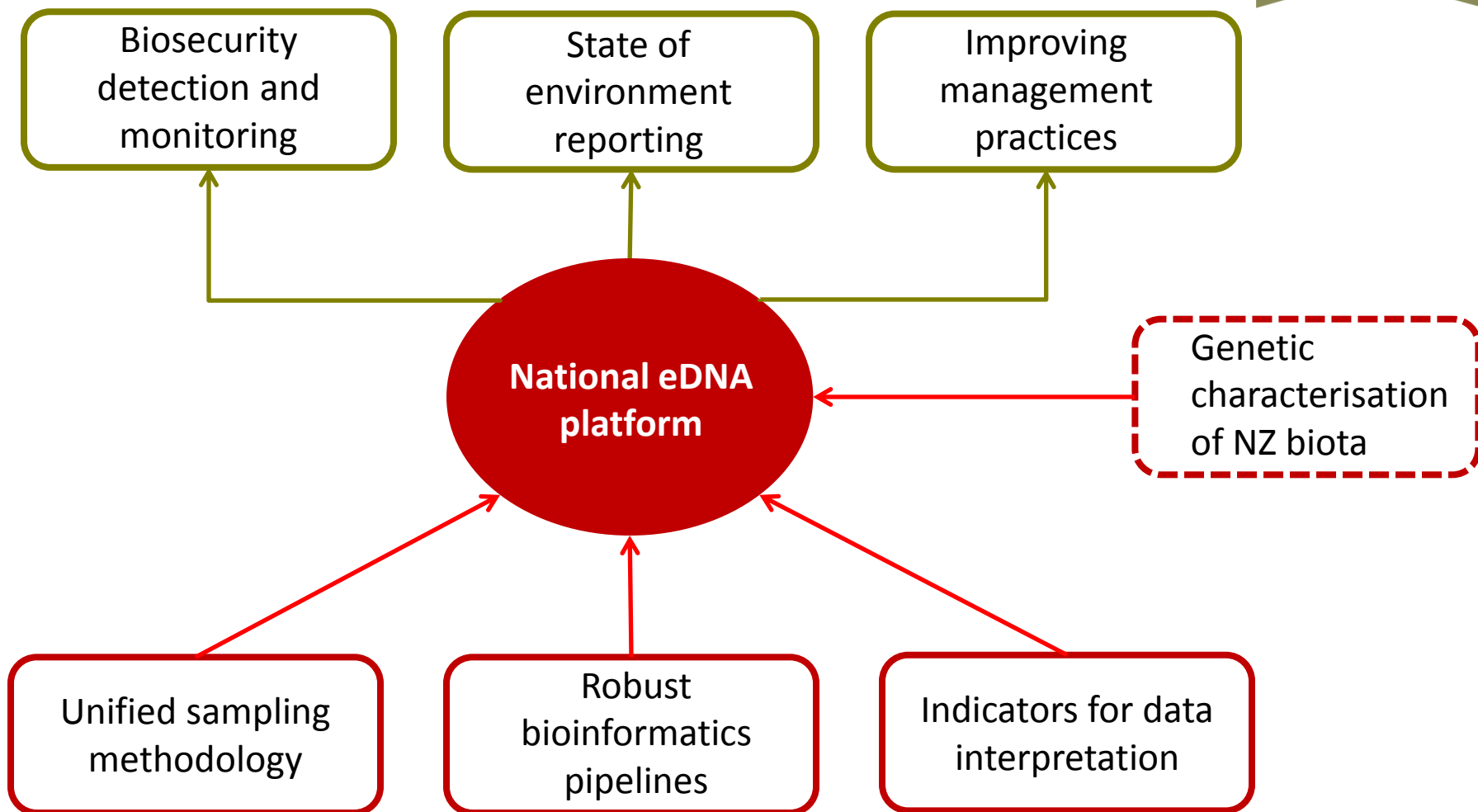
# A national framework for bioheritage assessment across natural and productive landscapes (Project 1.3)

- Aims to develop a New Zealand-wide framework for biodiversity assessment using environmental DNA data

## Two key focus areas

- Develop methods and infrastructure to realise the full potential of eDNA technology
- Build partnerships to consistently apply eDNA technology across the whole New Zealand landscape

# 5 year project outcome





# eDNA end-user workshop

Feeling inspired & thinking of potential eDNA applications?  
We want you to help shape the future of environmental DNA  
research in New Zealand

**When:** September 28<sup>th</sup>-29<sup>th</sup> 2015

**Where:** Wellington

Register your interest now

Robert Holdaway - [holdawayr@landcareresearch.co.nz](mailto:holdawayr@landcareresearch.co.nz)

Kevin Collins - [kcollinsconsult@gmail.com](mailto:kcollinsconsult@gmail.com)





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