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Fauna of New Zealand
Ko te Aitanga Pepeke o Aotearoa

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Periegopidae
(Arachnida: Araneae)

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Lincoln, Canterbury, New Zealand
2013
Class Periegopidae
Order Arachnida
Family Araneae

Periegopid spiders
There are only three species known in the family Periegopidae and all are in one genus, Periegops. These rare spiders have only ever been found in relict forests at limited locations in New Zealand (Banks Peninsula, Riccarton Bush, the Aldermen Islands, and East Cape) and Queensland, Australia. Periegopids are only found in forest with a deep leaf litter layer and well-drained soil. They do not build a web, but hunt on the forest floor. Periegopids can be most readily distinguished from other spiders found in New Zealand by having six eyes arranged in three widely spaced diads.

Contributor Cor Vink was born and educated in Christchurch, New Zealand. He completed a Ph.D. at Lincoln University on the taxonomy and systematics of New Zealand Lycosidae, a major part of which was published as a revision in *Fauna of New Zealand* 44. After completing his thesis he spent nine months at AgResearch as a postdoctoral research fellow investigating the genetics of hymenopteran parasitoids of weevil pests. From 2003 to 2005, Cor was a postdoctoral associate at San Diego State University, U.S.A., where he worked on developing new molecular markers for inferring deep phylogenetic relationships in spiders. At the end of 2005 he returned to New Zealand and joined the Biosecurity Group at AgResearch, Lincoln as a scientist. Cor is especially interested in the systematics of New Zealand spiders and is the adjunct curator of spiders at the Entomology Research Museum at Lincoln University. In 2013, Cor was appointed as Curator Natural History at Canterbury Museum, Christchurch.

(continued overleaf)

Ngā pūngāwere Periegopid
E toru anake ngā momo e mōhioitia ana o te whānau Periegopidae, ā, nō te huanga kotahi, arā, a Periegops. Kua kitea ēnei pūngāwere onge tonu i ngā toenga ngahere i ngā wāhi rauru nei i Aotearoa (i Horomaka, i te ngahere o Riccarton, i ngā moutere Aldermen, me Te Koringa-a-Paoa) me Queensland, i Ahitereiria. Kitea ai ēnei Periegopid i te ngahere he hōhonu nei ōna pūranga rau-rākau, i ngā one āhua pai te mimītanga wai. Kāore e hanga tukutuku mō rātou, engari ka whakangau i ā rātou kai i te papa tonu o te ngahere. Ka mōhioitia wawetia te Periegopid, ina whakaritea ki ētahi katoa o ngā pūngāwere o Aotearoa, nā te noho mai o ngā karu e ono, he tawhiti tonu te takoto o ia pūruatanga karu i ētahi atu.

Translation by Piripi Walker
Whakatiki
Contributor Nadine Dupérré is from Québec, Canada. She completed a Bachelor of Science at Université de Montréal in 1997. She began illustrating spiders in 1998 and to date has produced over 5000 illustrations. Nadine is probably best known amongst arachnologists for her illustrations in the book “Spiders of North America: an Identification Manual”. Nadine has published on the taxonomy of spiders and has also produced illustrations for publications on beetles and harvestmen. Nadine was a research assistant at the American Museum of Natural History, New York from 2008–2012 where she worked on Oonopidae with Norman Platnick. She is now living in Quito, Ecuador, working on a revision of the genus Agyneta (Linyphiidae) from North America, and is planning to continue her work on Linyphiidae in South America.

Contributor Jagoba Malumbres-Olarte is from the Basque Country, Spain. He completed a Ph.D. at Lincoln University on the ecology and diversity of spider communities in New Zealand native tussock grasslands in 2011. After he completed his thesis, he worked as a teaching assistant at Lincoln University and Imperial College London, UK. In the last six years Jagoba has been involved in numerous research projects on diverse topics, including the ecology of spider communities in sand dunes, conservation genetics of the Chatham Islands cockle weevil, the systematics of the New Zealand Clubiona, and the invasibility of spiders and plants in New Zealand. Jagoba is particularly interested in ecological and environmental processes that drive speciation and shape the diversity and composition of communities, and is currently developing new projects in this field.
ABSTRACT

Two species of Periegopidae, both in the genus *Periegops* Simon, 1893, are found in New Zealand; *P. suterii* (Urquhart, 1892) and *P. keani* sp. nov. The genus and both species are described or redescribed, with information on synonymy, type data, material examined, and geographical distribution. Habitus images of adults, illustrations of important morphological features, and distribution maps are provided. A key is given.

The mitochondrial gene cytochrome *c* oxidase subunit 1 (COI) and the nuclear gene 28S ribosomal RNA were sequenced for both species. COI divergence between some specimens of *P. suterii* was unexpectedly high (8.6%, uncorrected distance), but there was no morphological differentiation or 28S sequence divergence between those specimens. A molecular phylogenetic analysis examining the relationships between eight specimens of *P. suterii* and three specimens of *P. keani* using COI data is presented.

Keywords. Arachnida, Araneae, Periegopidae, New Zealand, Haplogynae, classification, distribution, ecology, biology, new species, key, phylogeny.


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CHECKLIST OF TAXA

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*suterii* (Urquhart, 1892)…………………………………….. 15

*keani* new species………………………………………… 17

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INTRODUCTION

Periegopidae Simon, 1893 is a family of six-eyed spiders comprising only three species, all in the genus *Periegops* Simon, 1893. Two species are endemic to New Zealand and the other is endemic to Queensland, Australia (Forster 1995). Periegopids can be distinguished from other families found in New Zealand by a combination of characters. The six eyes are arranged in three, widely spaced diads. The chelicerae have a lamina on the ventral surface and the maxillae are slender and are more than twice as long as wide and directed across the labium. The anterior two pairs of legs have asymmetrical superior claws; the proclaw of legs I and II has a double row of teeth whereas the retroclaw has a single row (Forster 1995: fig. 11–13). There is also a field of spicules on the median surface of the posterior median spinnerets (Labarque & Ramírez 2012; fig. 21F, 22E) and the venom outlet is anterior on the fang (Labarque & Ramírez 2012; fig. 18A); both of these characters require high microscope magnification to see them.

*Periegops suterii* (Urquhart, 1892) was first described from a female and two immature males collected by the early New Zealand zoologist, Henry Suter, at Dyer’s Pass on the Port Hills near Christchurch (Urquhart 1892; Forster 1995). Urquhart placed his new species in a Northern Hemisphere genus and family; *Segestria* Latreille, 1804, and *Dysderidae* C.L. Koch, 1837, respectively. Shortly after, Simon (1893) erected the genus *Periegops* for his new species *P. hirsutus*, which he described from a female specimen sent to him at the Muséum national d’Histoire naturelle, Paris, from New Zealand. Although Simon (1893) did not specify the location where the specimen was collected from, it was likely that it was from Banks Peninsula (Forster 1995). The generic description was of a female and Simon (1893: 267) noted that the male was unknown; however, the description of *P. hirsutus* was listed as male although no male characteristics appear to be described (Simon 1893: 268). Bryant (1935a: 54) and Forster (1995: 92) noted that the listing as a male appears to be in error. Dalmas (1917: 338) also noted the specimen was female and this was the specimen seen by Chamberlain (1946). It is unlikely that Simon realised he had described the same species as Urquhart and *P. hirsutus* was later made a synonym of *P. suterii* by Chamberlain (1946). Simon (1893) was first to recognise that the genus *Periegops* was distinct from other genera and placed it in its own subfamily, *Periegopinae* Simon, 1893. Forster (1995) elevated the subfamily to family status when he described *Periegops australis* Forster, 1995 and redescribed *P. suterii*. Forster (1995: 96) also noted that he had examined a single
female specimen collected at East Cape, but there were “no clear characters by which the species could be satisfactorily defined”; a male would need to be collected and the palpal bulb examined to determine if it was a separate species. Forster & Forster (1999) mentioned an additional specimen from the Aldermen Islands, which they believed to be a female, and Vink (2006) reported that he had sequence data from a fragment of the mitochondrial gene cytochrome c oxidase subunit 1 (COI) [specimen Pk1] that suggested that the North Island specimens were a distinct species.

Periegopidae is part of a group of spiders called the Haplogygae Simon, 1893, which are araneomorph spiders that lack separate fertilisation ducts, do not have a sclerotised epigyne and the male pedipalp is relatively simple. Haplogygae are a monophyletic group (Platnick et al. 1991; Ramírez 2000) that includes 17 families. Periegopidae have a number of characteristics that are shared with the haplogyne families Duguetiidae F. O. Pickard-Cambridge, 1899, Drymusidae Simon, 1893, Plectreuridae Simon, 1893, Sicytidae Blackwall, 1864, and Sicariidae Keyserling, 1880. These include a lamina on the ventral surface of the chelicerae, slender maxillary lobes directed across the labium, and a limited posterior respiratory system with fused apodemes (Forster 1995). Forster (1995) grouped this set of families, along with Periegopidae, into the superfamly Sicarioidea, but did not provide a formal definition of the superfamily. The name Sicarioidea was first used by Berland (1932) and Forster (1995) appears to be the only other arachnologist that has used it.

A more commonly used superfamily name that includes this set of taxa is Scytodoidea (Bristowe 1938; Caporiacco 1938; Brignon 1978; Lehtinen 1986); however, Scytodoidea has been used to refer to a larger set of haplogyne families that has also included Caponiidae Simon, 1890, Leptonetidae Simon, 1890, Ochyroceratidae Fage, 1912, Pholcidae C. L. Koch, 1850, and Tetrablemmidae O. Pickard-Cambridge, 1873. A morphological phylogenetic analysis by Platnick et al. (1991) supports a clade informally named “scytodooids” (Coddington & Levi 1991) that includes Duguetiidae, Drymusidae, Leptonetidae, Ochyroceratidae, Pholcidae, Plectreuridae, Sicytidae, Sicariidae, Telemidae Fage, 1913, and Tetrablemmidae. The monophyly of this clade is supported by the reduction of the posterior spiracles to one and the tetrahedral posterior median spinnerets (Platnick et al. 1991), although these characters have been reversed in some species in the clade (Platnick et al. 1991).

Ramírez (2000) examined the morphology of haplogyne respiratory systems and added characters to the data matrix of Platnick et al. (1991) to produce a phylogeny of the Haplogygae. Periegopidae was not included in the analyses but Ramírez (2000) stated that Periegopidae was sister to the Scytodidae. Periegopidae was also included in the scytodoid clade by Coddington (2005); its inclusion was presumably based on Forster (1995) and Ramírez (2000). Labarque & Ramírez (2007a, b) considered that among the scytodoids, Periegopidae was closest to Drymusidae and Scytodidae as they share two characters; asymmetrical superior claws on the anterior two pairs of legs and a field of spicules on the median surface of the posterior median spinnerets. More recently, Labarque & Ramírez (2012) expanded the morphological dataset of Platnick et al. (1991) and Ramírez (2000) to include Periegopidae along with a restricted Scytodoidea, which included Drymusidae, Scytodidae, and Sicariidae. Periegops, Drymusia, and Sicytidae formed a clade, which all shared the synapomorphies of asymmetrical superior claws and a field of spicules on the median surface of the posterior median spinnerets. Labarque & Ramírez (2012) concluded that Periegopidae was sister to Drymusidae, but they did not find any autapomorphies for Periegops and suggested that the two families might be synonymised.

Very little is known of the biology of periegopids. They are only found in forest with a deep leaf litter layer and well-drained soil. Specimens have been collected from under logs and rocks, in leaf litter, in pitfall traps, in rotted logs and root cavities, and in a mygalomorph burrow (Forster 1995; Vink 2006). Forster (1995) suggested that periegopids were rare as their habitat has been severely reduced due to human deforestation and only 21 Periegops specimens had been collected in New Zealand and Australia between 1891 and 1995. In surveys between October 2002 and May 2003, Vink (2006) found a further 24 specimens of P. suterii at ten different locations on the Banks Peninsula. Sirvid et al. (2012) classified P. suterii as having a relict distribution.

Periegops do not appear to build a web for prey capture, but do build silken retreats. It seems probable that periegopids are cursorial, night-time hunters as they have only been found on the forest floor, have been collected in pitfall traps and lack webs. On two occasions a single female has been found together with two or three males under logs and rocks (Forster 1995; Vink 2006), which implies that the female might possess some method of attracting males.

MORPHOLOGY AND TERMINOLOGY

The morphological structures required for the identification of New Zealand and Australian Periegopidae are
referred to in Fig. 1–16 and explained in the glossary of technical terms (Appendix A), and Paquin et al. (2010). As with almost all spiders, the male pedipalp is important when identifying periegopids to species. The female epigyne, which is usually useful for identifying spider species (Paquin et al. 2010), is just a lightly sclerotised area that covers the internal genitalia in periegopids, and many other haplogyne spiders, and therefore is not likely to be diagnostic. The internal genitalia, however, are diagnostic, as is the case with most spider species. The morphological nomenclature of the pedipalp and the internal genitalia follows Forster (1995).

A phylogenetic species concept (Wheeler & Platnick 2000) has been implemented in this study. It defines a species as the smallest aggregation of populations diagnosable by a unique combination of character states.

**METHODS AND CONVENTIONS**

**Collecting.** Periegopids can be collected by a variety of methods. The best method for collecting periegopids is by searching under rocks and logs that are sitting on or partially embedded in soil on open ground within forest in areas with a good litter layer over well drained soil (Vink 2006). Periegopids have also been found under wooden discs (M. H. Bowie, pers. comm.) that are facsimiles for natural fallen logs (Bowie & Frampton 2004). Periegopids have also been collected by sieving and searching leaf litter. Specimens of *P. suterii* and *P. australis* have been caught in pitfall traps, but unless specimens are collected and preserved within a couple of days of being caught they can start to decay, which can make identification difficult. Decay of specimens caught in pitfall traps can be mitigated by the use of a good preservative such as propylene glycol, which also preserves DNA but can shrivel soft tissue (Vink et al. 2005). Malumbres-Olarte et al. (in press) used propylene glycol in their pitfall traps and found that useful DNA was preserved after two weeks in the field.

**Preservation.** Periegopids are best preserved long-term in 70–75% ethanol. To ensure adequate DNA preservation, specimens should also be stored at ≤ -20°C (Vink et al. 2005). Spiders can be stored in 95–100% ethanol to preserve DNA but as with lower ethanol concentrations, it is still best to combine this with storage temperatures ≤ -20°C (Vink et al. 2005). Preservation in 95–100% ethanol makes specimens brittle, potentially rendering them unsuitable for morphological examination.

**Preparation.** Specimens should be labelled with the locality, including area code (Crosby et al. 1976, 1998), latitude and longitude, collection date, collector’s name, habitat data, and collection method.

Most morphological features used in identification can be seen under an ordinary dissecting microscope. When examining a spider in ethanol it should be rested in washed quartz sand or glass beads to provide support for the specimen. This also allows the specimen to be positioned at any desired viewing angle. The features of the male pedipalp are best viewed by removing the left pedipalp at the junction between the trochanter and the femur, and viewed ventrally. In periegopids, as with other haplogyne spiders, there is no sclerotised external genitalia, only a slightly sclerotised plate, which makes distinguishing females from immature specimens difficult. Female internal genitalia were excised using a sharp entomological needle and were prepared for examination by placing the dissected genitalia in either lactic acid or 10% KOH solution for 1–3 hours at 37°C to dissolve soft tissue. Internal genitalia were only illustrated for *P. suterii* and *P. australis*.

**Measurements.** All measurements are in millimetres (mm). Where the measurements are expressed as a fraction, the numerator refers to the length of the structure and the denominator refers to its width. Measurements outside parentheses are for males and inside parentheses for females. The size ranges given for the body and carapace length of each species represent the smallest and largest individuals of each sex that we examined. The mean body length and carapace length were calculated and the number of specimens measured given.

**Descriptions.** For the new species, illustrations, measurements and colour pattern descriptions were made from the type specimen. For the existing species, illustrations, measurements, and colour pattern descriptions were prepared from a non-type representative male and female specimen (with collection information shown).

Descriptions of colours are for ethanol-preserved specimens. It should be noted that colours and colour patterns can fade in older specimens, particularly those that have not been stored away from light.

Measurements were made with a micrometer ruler fitted to the eyepiece of a Leica M125 stereo microscope. Characters diagnostic in other spider families (e.g., eye size and position, leg spination) are not diagnostic for Periegopidae species and have not been included in the descriptions.
Illustrations. Specimens to be illustrated were first photographed with a Nikon Coolpix 950 digital camera attached to a SMZ-U Nikon dissection microscope. The digital photos were then used to establish proportions and the illustrations were detailed and shaded by referring back to the structure under the microscope.

Map images were created using the geographic information system software ArcMap 10 (ESRI).

Text conventions. The area codes of Crosby et al. (1976, 1998) are used in collection records.

The following acronyms for repositories are used:

- AMNH American Museum of Natural History, New York, U.S.A.
- LUNZ Entomology Research Museum, Lincoln University, New Zealand
- MONZ Museum of New Zealand Te Papa Tongarewa, Wellington, New Zealand
- NZAC New Zealand Arthropod Collection, Auckland, New Zealand
- OMNZ Otago Museum, Dunedin, New Zealand
- QMB Queensland Museum, Brisbane, Australia

Molecular biology. To construct a molecular phylogeny of New Zealand Periegopidae and to facilitate the identification of immature specimens, we used the mitochondrial gene cytochrome c oxidase subunit 1 (COI) and the nuclear ribosomal RNA gene 28S. COI is one of the fastest evolving mtDNA genes and has been used to examine genetic differences between and among haplogyne spiders (e.g., Arnedo et al. 2001, 2009; Astrin et al. 2006; Starrett & Waters 2007; Binford et al. 2008; Dimitrov et al. 2009; Huber & Astrin 2009; Duncan et al. 2010) and New Zealand spider species (Vink & Patterson 2003; Vink et al. 2008, 2011a, b; Framenau et al. 2010; Vink & Dupéré 2010; Lattimore et al. 2011; Malumbres-Olarte & Vink 2012). 28S was selected because it is a slow evolving gene (Hedin & Maddison 2001) and has been used in phylogenetic analyses of haplogynes (Bruvo-Madaric et al. 2005; Binford et al. 2008) as well as other spiders (e.g., Hedin & Bond 2006; Rix et al. 2008; Wang et al. 2008; Spagna et al. 2010). Eight specimens of P. suterii and three specimens of P. keani were sequenced for COI. Each specimen was arbitrarily assigned a specimen code (Table 1). A subset of four specimens, two from each species, was sequenced for 28S.

DNA was extracted non-destructively (see Paquin & Vink 2009) from either two to three legs using a ZR Genomic DNA™ Tissue Minipreps (Zymo Research). The primers used to PCR amplify and sequence COI fragments were either LCO1490 (5'GGAATTTGAGATATTGGC') (Folmer et al. 1994) plus C1-N-2776-spider (5'GGATAATT-CAGAATANCGGAG-3') (Vink et al. 2005) or C1-J-1718-spider (5'GGGAGTTTGGAAATGGRT-TRRTCC-3') (Vink et al. 2005) plus C1-N-2776-spider. The two COI fragments were 1260 base pairs (bp) and 1055 bp long, respectively. Two different primer pairs were used to amplify and sequence two overlapping 28S fragments; 28S-B1 (5'GACCCGATAGCAAAACATACCCG-3') (Bruvo-Madaric et al. 2005) plus 28S-B2 (5'GATTAGTCTTCGCCCTTATA-3') (Bruvo-Madaric et al. 2005) and 28Sa (5'GACCCGTCTTGAACACCGA-3') (Nunn et al. 1996) plus LSUR (5'GCTACTCACCAAGATGCTGCA-3') (Rix et al. 2008). The two 28S fragments were 819–821 bp and 830–833 bp long, respectively. PCR amplification was performed using i-StarTaq™ DNA Polymerase (iNtRON Biotechnology) in a Mastercycler® (Eppendorf) thermocycler with a cycling profile of 35 cycles of 94°C denaturation (30 s), 48°C (COI) or 60°C (28S) annealing (30 s), 72°C extension (1 min) with an initial denaturation of 3 min and a final extension of 5 min. Excess primers and salts were removed from the resulting double-stranded DNA using a DNA Clean & Concentrator™ Kit (Zymo Research). Double bands were observed when 28S PCR products of Pk1 were visualised via gel electrophoresis; both bands were excised from the gel and prepared for sequencing using a Zymoclean™ Gel DNA Recovery kit. For Pk1, the primer pair 28S-B1 plus 28S-B2 amplified a fragment of 28S DNA that BLAST database searching indicated may have come from a ciliate protozoan. For Pk2, the primer pair 28Sa plus LSUR amplified a fragment of 28S DNA from a soil nematode in the family Cephalobidae. Amplification of 28S was attempted for specimens Ps5 and Ps6, but was not possible owing to contamination by a fungus that the primers preferentially annealed to. Contaminants were identified by BLAST searching their sequences (Altschul et al. 1997). Purified PCR fragments were sequenced in both directions at the Core Instrumentation Facility (University of California, Riverside, USA), the Massey Genome Service (Massey University, New Zealand), or Macrogen (Korea). Sequence data were deposited in GenBank (www.ncbi.nlm.nih.gov/GenBank/ – see Table 1 for accession numbers). Sequences were edited using Sequencher 4.6 (Gene Codes Corporation).
### Table 1. Specimens used for molecular analysis.

<table>
<thead>
<tr>
<th>Species</th>
<th>Specimen code</th>
<th>Sex</th>
<th>Collection information</th>
<th>GenBank accession numbers</th>
</tr>
</thead>
<tbody>
<tr>
<td><em>Periegops suterii</em></td>
<td>Ps1</td>
<td>♂</td>
<td>MC, Hinewai Reserve, 43°48.59'S, 173°01.28'E, 11 October 2002, CJV, AMNH (ARAMR-000615)</td>
<td>JX174281</td>
</tr>
<tr>
<td><em>Periegops suterii</em></td>
<td>Ps2</td>
<td>♀</td>
<td>MC, Hinewai Reserve, 43°48.59'S, 173°01.28'E, 11 October 2002, CJV, AMNH (ARAMR-000616)</td>
<td>JX174282</td>
</tr>
<tr>
<td><em>Periegops suterii</em></td>
<td>Ps3</td>
<td>♀</td>
<td>MC, Hay Reserve, 43°42.18'S, 172°53.87'E, 13 March 2003, CJV, LUNZ (00012720)</td>
<td>JX174283</td>
</tr>
<tr>
<td><em>Periegops suterii</em></td>
<td>Ps4</td>
<td>♂</td>
<td>MC, Kennedys Bush Reserve, 43°37.9'S, 172°37.3'E, 22 April 2011, CJV, H.P. &amp; L.J. Hudson Vink, LUNZ (00012715)</td>
<td>JX017357, JX017353</td>
</tr>
<tr>
<td><em>Periegops suterii</em></td>
<td>Ps5</td>
<td>♂</td>
<td>MC, Montgomery Park Reserve, 43°44.7'S, 172°52.2'E, 5 November 2010, CJV &amp; JM-O, LUNZ (00012722)</td>
<td>JX174284</td>
</tr>
<tr>
<td><em>Periegops suterii</em></td>
<td>Ps6</td>
<td>♂</td>
<td>MC, Hinewai Reserve, 43°48.69'S, 173°00.95'E, 11 January 2011, M.H. Bowie, LUNZ (00012735)</td>
<td>JX174285</td>
</tr>
<tr>
<td><em>Periegops suterii</em></td>
<td>Ps7</td>
<td>subadult ♀</td>
<td>MC, Montgomery Park Reserve, 43°44.7'S, 172°52.2'E, 5 November 2010, JM-O &amp; CJV, LUNZ (00012725)</td>
<td>JX174286, JX174291</td>
</tr>
<tr>
<td><em>Periegops suterii</em></td>
<td>Ps8</td>
<td>♀</td>
<td>MC, Kennedys Bush Reserve, 43°37.9'S, 172°37.3'E, 13 August 2011, CJV &amp; H.P. Hudson Vink, LUNZ (00012718)</td>
<td>JX174287</td>
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<tr>
<td><em>Periegops keani</em></td>
<td>Pk1</td>
<td>immature</td>
<td>CL, Ruamahuanui Island, 36°57.25'S, 176°05.52'E, 19 November 2003, B.M. Fitzgerald, MONZ (AS.002310)</td>
<td>JX174288, JX174292</td>
</tr>
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<td><em>Periegops keani</em></td>
<td>Pk2</td>
<td>♂</td>
<td>CL, Ruamahuanui Island, 36°57.25'S, 176°05.52'E, 23 March 2012, CJV &amp; J.M. Kean, LUNZ (00012716)</td>
<td>JX174289, JX174293</td>
</tr>
<tr>
<td><em>Periegops keani</em></td>
<td>Pk3</td>
<td>immature</td>
<td>CL, Ruamahuanui Island, 36°57.25'S, 176°05.52'E, 23 March 2012, CJV &amp; J.M. Kean, LUNZ (00012717)</td>
<td>JX174290</td>
</tr>
</tbody>
</table>
PHYLOGENETIC ANALYSIS

Methods. Sequences were aligned using Sequencher 4.6. There was no evidence of insertions/deletions or stop codons in the COI sequences and alignment was straightforward.

Uncorrected COI pairwise distances were calculated using PAUP* version 4.0b10 (Swofford 2002). Distances using the Kimura-2-Parameter (K2P) model (Kimura 1980) were also calculated for comparison with previously reported intraspecific distances in spiders (Robinson et al. 2009). Although K2P has been commonly used in most DNA barcoding studies (e.g., Robinson et al. 2009), there is no evidence that this model is better for species identification than simpler metrics such as uncorrected pairwise distances (Astrin et al. 2006; Collins et al. 2012).

Partitioned Bayesian analysis implemented in MrBayes version 3.1.2 (Ronquist & Huelsenbeck 2003) was used to estimate the COI phylogenetic tree topology. MrModeltest version 2.3 (Nylander 2008) implemented in PAUP* version 4.0b10 (Swofford 2002) was used to select the optimal model and model parameters. Within MrModeltest the Akaike Information Criterion was used for model selection (Posada & Buckley 2004). Based on the results of Brandley et al. (2005), the COI data were partitioned by codon with models selected for each codon; GTR+I (Lanave et al. 1984; Tavaré 1986) for the 1st and 3rd codon positions and HKY+I (Hasegawa et al. 1985) for the 2nd codon positions. Bayesian analysis was conducted by running two simultaneous, completely independent analyses each with four heated chains, sampling every 1000th tree. The analysis was run for 20 million generations, by which time the average standard deviation of split frequencies had dropped below 0.002, which indicated that the two tree samples had converged. Tracer version 1.5 (Rambaut & Drummond 2009) was also used to determine if the analyses had sufficient effective sample sizes. MrBayes was used to construct majority rule consensus trees, discarding the first 25% of trees generated as burn-in. TreeView 1.6.6 (Page 1996) was used to view and save trees in graphic format.

Results. A 1031 base pair (bp) COI fragment was sequenced from eight specimens of P. suterii and three specimens of P. keani. Four COI haplotypes occurred among the eight specimens of P. suterii. Each of the three specimens of P. keani had a different COI haplotype. Inter- and intraspecific uncorrected pairwise distances between COI sequences of P. suterii and P. keani are shown in Table 2. The minimum divergence between the two species was 13.0% (uncorrected) and the mean divergence was 13.5% (uncorrected). The mean intraspecific divergences in P. suterii and P. keani, were 4.0% (uncorrected) and 0.6% (uncorrected), respectively. The maximum intraspecific divergence in P. suterii was 8.6% (uncorrected, 9.2% K2P-distance) and was 0.7% (uncorrected and K2P-distance) in P. keani.

The 1360 bp fragments of 28S from two specimens of P. suterii were identical. The amplification of 28S from two specimens of P. keani (Pk1 and Pk2) was only partially successful with an 821 bp fragment from Pk1 and an 833 bp fragment from Pk2; however, the 281 bp overlap was identical. There were 12 nucleotides that varied between the two species.

The phylogenetic analysis of the COI data (Text-fig. 1) showed that P. suterii and P. keani are monophyletic with P. suterii divided into two clades.

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Discussion. *Periegops* COI sequences were very similar or identical (Table 2) for specimens collected at the same sites; however, specimens of *P. suterii* collected at Kennedys Bush Reserve, in the west of the Banks Peninsula (Map 1), were 8.2–8.6% (8.7–9.2% K2P-distance) divergent from *P. suterii* specimens collected from three localities in the eastern Bank Peninsula (Map 1). This high divergence was not found in the slower evolving nuclear marker; 28S sequences were identical from specimens from Kennedys Bush Reserve and Montgomery Park Reserve (Map 1). We also did not observe any morphological differences between Kennedys Bush Reserve specimens and specimens collected from other locations on the Banks Peninsula; therefore we are confident that all specimens are one species, *P. suterii*. A difference of 8.7–9.2% (K2P-distance) in COI sequence divergence is very high in spiders; much more than the average maximum intraspecific divergence of 3.16% (K2P-distance) observed in other spiders (Robinson et al. 2009). However, higher intraspecific divergences (>10%) have been observed in other haplogynes (Astrin et al. 2006; Binford et al. 2008), Hypochilidae (Hedin 2001), and Mygalomorphae (Bond et al. 2001), so perhaps COI divergence is higher in ancestral spiders than it is in the Entelegynae. Nevertheless, it is curious that high (8.2–8.6%) COI divergence was observed between specimens from localities only just over 23 km apart and at similar altitudes, while there was much less divergence (0.7–1.3%) between specimens from eastern Bank Peninsula localities, which were 5–15 km apart. Specimens from localities between Kennedys Bush Reserve and Montgomery Park Reserve, such as Kaituna Valley, could reveal if COI divergence is correlated with distance or whether there has been some sort of genetic isolation between the east and west of Banks Peninsula, similar to the geographic distribution of two closely related weta species in the genus *Hemideina* (Townsend et al. 1997).

Text-fig. 1 Unrooted Bayesian consensus tree based on cytochrome c oxidase subunit I (COI) sequence data. Values on branches are posterior probabilities. Specimen codes are listed in Table 1. Branch lengths are proportional to the expected number of substitutions per site (see scale bar).
KEY TO NEW ZEALAND PERIEGOPIDAE

1 Tibia of leg I less than 0.7× length of carapace. When viewed retrolaterally, dorsal surface of cymbium of male pedipalp projects dorsally (Fig. 9). Flared flattened flanges near embolus tip (Fig. 6, 7). Found only on Banks Peninsula and Riccarton Bush (Map 1). ........................ (p. 15) Periegops suterii (Urquhart)

—Tibia of leg I more than 0.75× length of carapace. When viewed retrolaterally, dorsal surface of cymbium of male only slightly projects dorsally (Fig. 9). Flattened flanges near embolus tip not flared (Fig. 9a). Found only on Aldermen Islands and East Cape (Map 2). ............... (p. 17) Periegops keani sp. nov.

BIOSYSTEMATICS

Family PERIEGOPIDAE

Medium-sized, 3-clawed haplogyne spiders. 6 eyes in 3 widely separated diads (Fig. 1, 14, 15). Proclaw of legs I and II with double row of teeth, retroclaw with single row. Chelicerae with lamina on the ventral surface (Fig. 2) and slender maxillary lobes directed across the labium (Fig. 3). Male pedipalp simple, consisting of a bulb with a short embolus (Fig. 6–9); coiled sperm duct often visible within the bulb. Abdomen ovoid, dun or dirty cream usually with a distinct chevron pattern (Fig. 1, 14–16) and black setae. Slightly sclerotised area anterior to the epigastric furrow (Fig. 10, 11). Female internal genitalia simple, haplogyne, consisting of a rounded poreplate with many small invaginated cups (Fig. 12, 13). 6 spinnerets and small colulus; median surface of posterior median spinnerets with field of spicules (Labarque & Ramírez 2012; fig. 21F, 22E).

Forster (1995) provided details of the respiratory system of Periegops.

Periegops suterii (Urquhart)

Fig. 1, 4, 6, 7, 10, 12, 14, 16; Map 1
Segestria suterii Urquhart, 1892: 230.

Diagnosis. Distinguished from all other Periegops species by the shape of the embolus and cymbium of the male pedipalp. Flattened flanges near embolus tip more pronounced than those of P. keani, and embolus is shorter than that of P. australis. Dorsal surface of cymbium projects slightly more than that of P. keani, but much less than that of P. australis. Poreplate of internal genitalia not as rounded as that of P. australis. Leg I of males, females, and immatures shorter than in P. keani; tibia I less than 0.7× length of carapace. Found only on Banks Peninsula and Riccarton Bush in the South Island.

Description. Carapace: red-orange anteriorly, orange posteriorly; with sparse black setae; fovea absent. Sternum: orange. Leg I orange-brown, patella, distal ends of femur and tibia light orange; legs II, III, and IV yellow-brown, darker at proximal end of each leg segment. Abdomen: dun with black setae; black-brown chevron pattern (Fig. 1, 14, 16). Chelicerae red-brown. Male pedipalp simple with a bulb and a short embolus, which is flattened and
twisted, flaring out to 2 flanges just before the tip (Fig. 6, 7); coiled sperm duct often visible within the bulb. Slightly sclerotised area anterior to epigastric furrow (Fig. 10). Female internal genitalia simple, haplogyne, consisting of a rounded poreplate with many small invaginated cups (Fig. 12).

**Dimensions** (mm). Male MC, Kennedys Bush Reserve LUNZ 00012715 (female MC, Kennedys Bush Reserve LUNZ 00012718): total length 6.13 (7.92); carapace length 3.06/2.05 (3.82/2.60), height 1.15 (1.71); abdomen 2.87/1.80 (4.12/2.60); sternum 1.64/1.05 (1.93/1.25).

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<th>IV</th>
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<td>(7.16)</td>
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The ratios of the length of tibia I to the length of the carapace (measured for all specimens, including immatures) were between 0.54 and 0.70. The ratio of leg I segment length to carapace length has also been shown to be a reliable character for the separation of other closely related spider species (Vink et al. 2008).

The chevron pattern on the abdomen (Fig. 1, 14, 16) was visible in all but one of the specimens examined, a male (LUNZ 00012715). Forster (1995) noted that a female collected at Riccarton Bush did not have abdominal markings.

**Type data.** Type of Segestria suterii: Not examined. Original description based on a female specimen from “Dyer’s Pass, Canterbury”, which was collected with 2 immature males by H. Suter (Urquhart 1892). Forster (1995) stated the type was in poor condition in the Canterbury Museum, Christchurch, New Zealand, where all of Urquhart’s existing type specimens are housed (Court & Forster 1988; Nicholls et al. 2000; Paquin et al. 2008). However, the type of Segestria suterii was not included in the list of arachnid primary types held in Canterbury Museum (Nicholls et al. 2000). It was not possible to check whether the type specimen was present at Canterbury Museum, as the collections have not been accessible since the Christchurch earthquake in February 2011 (S. D. Pollard, pers. comm.). The type is not in OMNZ (C. Fraser, pers. comm.), which houses New Zealand’s largest collection of spiders. It is possible that the type was amongst specimens at Ray Forster’s house at the time of his death and has as yet not been located. We consider the type of Segestria suterii missing, but not lost.

**Type of Periegops hirsutus:** Not examined. Original description based on a female specimen from an undisclosed location in New Zealand (Simon 1893). Although many of Simon’s type specimens are housed in the Muséum national d’Histoire naturelle, Paris, France, the type of Periegops hirsutus could not be found there (C. Rollard, pers. comm.).

**Material examined.** 32 non-type specimens (15 males, 15 females, 2 subadult females) — see Appendix B for collection details of specimens examined.

**Distribution** (Map 1). Found only on Banks Peninsula and Riccarton Bush (MC), Forster (1995) and Vink (2006) listed localities where P. suterii specimens have been found. Riccarton Bush is the only site outside of the Banks Peninsula where P. suterii has been found (in 1994), but subsequent searches there (Chinn 2006; Vink 2006) have failed to find it.

**Biology.** Periegops suterii has been found in forest with a deep leaf litter layer and well-drained soil. It has been found under logs and rocks, and in leaf litter, in both beech and podocarp forest. Within suitable locations, P. suterii has a patchy distribution, but specimens are often found years apart in the same specific locations. Periegops suterii does not appear to build a web for prey capture, but it does produce dragline silk and builds silken retreats. The lack of webs and their presence in pitfall traps suggest that it is a cursorial, night-time hunter. On two occasions a single female has been found together with two or three males under logs and rocks (Forster 1995; Vink 2006), which implies that the female might possess some method of attracting males. Forster (1995) suggested pheromones, but it could also be stridulation; Forster (1995) did not notice the stridulatory ridges on the chelicerae. Adults have been found from August to May.

**DNA.** Nine cytochrome c oxidase subunit I and two 28S rRNA sequences for this species are listed in GenBank (www.ncbi.nlm.nih.gov/GenBank/) under accession numbers EF537066, JX017353, JX017357, JX174281–JX174287, and JX174291.
Remarks. Although the types of Segestria suterii and Periegops hirsutus were not examined, we have no reason to doubt that these species are synonymous, as Chamberlain (1946) examined the type specimens of both species. Forster (1995: 92) stated that it can “be reliably assumed” that the type of P. hirsutus “came from the same locality as Urquhart’s Segestria suterii.” Also, Simon (1893) noted that the type specimen of P. hirsutus was 8 mm long, which is within the range of female body lengths we observed for P. suterii.

Despite the common usage of the name Periegops suteri, the correct original spelling of the specific epithet is suterii (as Segestria suterii Urquhart, 1892). Article 33.4 in the fourth edition of the International Code of Zoological Nomenclature (International Commission on Zoological Nomenclature 1999) states the change from -i to -ii is an incorrect subsequent spelling. There is provision in the International Code of Zoological Nomenclature for prevailing usage of an unjustified emendation (article 33.2.3.1) or incorrect subsequent spelling (article 33.3.1); however, suteri is not an emendation but an incorrect subsequent spelling and article 33.4 is an exception to articles 33.2.3.1 and 33.3.1. Both spellings have been used: P. suteri by Forster (1995), Forster & Forster (1999), Paquin et al. (2010), and Sirvid et al. (2011, 2012), and P. suterii by Bryant (1935a, b), Chamberlain (1946), Forster (1967), Forster & Forster (1973), Vink (2006), Jocqué & Dippenaar-Schoeman (2006), Brockerhoff et al. (2008), Platnick (2012), and Labarque & Ramírez (2012). As this is not a case of prevailing usage of an incorrect subsequent spelling (article 33.3.1) and article 33.4 is quite clear on the matter, we advocate for and use the correct original spelling.

Periegops keani new species
Fig. 2, 3, 5, 9, 15; Map 2

Diagnosis. Distinguished from all other Periegops species by shape of embolus and cymbium of the male pedipalp. Flattened flanges near embolus tip not as pronounced as those of P. suterii and embolus shorter than that of P. australis. Dorsal surface of cymbium projects only slightly, less than that of P. suterii and much less than that of P. australis. Leg I of male and immatures longer than in P. suterii; tibia I more than 0.75× length of carapace. Found only on the Aldermen Islands and East Cape in the North Island.

Description. Carapace: dark orange-brown anteriorly, orange-brown posteriorly; with sparse black setae; fovea absent. Sternum: light orange brown. Leg I dark orange-brown, patella, distal ends of femur, and tibia orange-brown; legs II, III, and IV orange-brown, darker at proximal end of each leg segment. Abdomen: dirty cream with black setae; chevron pattern barely visible (Fig. 15). Chelicerae red-brown. Male pedipalp simple with a bulb and a short embolus, which is flattened and twisted, flaring out slightly to 2 flanges just before the tip (Fig. 9); coiled sperm duct visible within the bulb. Female unknown.

Dimensions (mm). Male holotype: total length 10.03; carapace 4.99/3.48, height 1.66; abdomen 5.25/3.75; sternum 2.40/1.90.

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*Both tarsi IV were damaged so the length was estimated based on the proportions observed in immature species of P. keani.

Variation. The holotype male was the only specimen that did not have abdominal markings; all of the other subadult and immature specimens had a chevron pattern on the abdomen, like that of P. suterii (Fig. 1, 14, 16).

The ratios of the length of tibia I to the length of the carapace (measured for all specimens, including immatures) were between 0.77 and 0.88. Based on what we observed for P. suterii, it seems likely that female P. keani will have a tibia/carapace ratio within that range.

Type data. Holotype: male (LUNZ 00012716) labelled “NEW ZEALAND, CL, Ruamahuanui Island; 36°57.25’S, 176°05.52’E; NW campsite, under rock; 23 Mar 2012, C.J. Vink & J.M. Kean” (LUNZ).

Material examined. Type specimen plus 7 non-type specimens (1 subadult male, 1 subadult female, 5 immatures) — see Appendix B for collection details of specimens examined.

Distribution (Map 2). Found only on the Aldermen Islands and East Cape (CL, GB).

Biology. Periegops keani has been found in forest with a deep leaf litter layer and well-drained soil. On the forest floor it has been found under rocks and in a Stanwellia sp. (Araneae: Nemesiidae) burrow (Forster 1995). Periegops keani does not appear to build a web for prey capture and
probably builds silken retreats, like *P. suterii*. The lack of webs suggests that it is a cursorial, night-time hunter. Adults have been found in March and September, but given that the climates it is found in are milder than those of *P. suterii*, it seems likely that adults occur throughout the year.

**DNA.** Three cytochrome *c* oxidase subunit 1 and two 28S rRNA sequences for this species are listed in GenBank (www.ncbi.nlm.nih.gov/GenBank/) under accession numbers JX174288–JX174290, JX174292, and JX174293.

**Etymology.** The specific name is in honour of the senior author’s friend and colleague, John Kean, who helped collect the type specimen.

**Remarks.** The bulb of the holotype was not oriented to the same angle with respect to the cymbium (Fig. 9) as the specimens illustrated of *P. suterii* (Fig. 6, 7) and *P. australis* (Fig. 8); however, when oriented in a slight dorsal direction (Fig. 9a), it looks the same as those of *P. suterii*, other than the flattened flanges near the embolus tip, which are not as pronounced in *P. keani*.

We consider that a female specimen found in forest below the East Cape Lighthouse is *P. keani*. Forster (1995) remarked that the genitalia of this specimen was indistinguishable from that of *P. suterii*. The female specimen from East Cape was collected by Grace Hall (NZAC) on 30 September 1993 and loaned to the late Ray Forster, but has not been returned to NZAC (G. Hall pers. comm.) and it has not been located at OMNZ (C. Fraser, pers. comm.). Unsuccessful searches for further specimens have been conducted in the small (~10 hectare) patch of forest below the East Cape Lighthouse in October 1994 (G. Hall), February 1995 (G. Hall), September 1995 (G. Hall & P. J. Sirvid), November 1995 (G. Hall & L. J. Boutin), December 2009 (G. Hall), November 2010 (J. Malumbres-Olarte) and February 2012 (C. J. Vink). Searches of the forest floor (including rock and log turning) were conducted in all trips, *Stanwellia* sp. burrows were searched in most trips, and pitfall trapping and litter searches were also conducted in the former and later trips. *Periegops keani* may conceivably now be extinct from that site.

It is possible that the population at East Cape could be a separate species to *P. keani*, as *Periegops* probably has limited dispersal ability. Although the Aldermen Islands and East Cape are 233 km apart, the Aldermen Islands would have been connected to the mainland from the Last Glacial Maximum (McKinnon 1997; Trewick & Bland 2012) up until 12000 years ago (Schofield & Thompson 1964; McKinnon 1997) and there would have been continuous forest between the two sites (McKinnon 1997; Trewick & Bland 2012). If a fresh specimen could be located at East Cape, 28S sequence data could confirm whether they were the same species.

Forster & Forster (1999) stated that they had examined a female specimen from the Aldermen Islands. This specimen was collected from Ruamahauti Island in November 1972 and loaned to Ray Forster from NZAC (G. Hall, pers. comm.). We have examined this specimen and determined it to be a subadult male, as it has swollen palpal tibiae, which are characteristic of males in their final moult before adulthood. The specimen did not have an epigyne or internal genitalia and we believe Forster & Forster (1999) made a typographical error in stating the specimen was female. We consider that this specimen is *P. keani*. Ruamahauti Island is only 2 km from Ruamahuaui Island and the land below the two islands is no more than 20 m under the sea (Hayward & Moore 1973), therefore, the two populations would have been connected up until 9000 years ago (Schofield & Thompson 1964; McKinnon 1997).

Size may help differentiate *P. keani* from *P. suterii* even though we have only examined a single adult specimen of the former species. The subadult male from Ruamahauti Island is larger (body length = 8.8 mm, carapace length = 4.3 mm) than any of the 11 adult male *P. suterii* that we have examined. Also, a *P. keani* specimen (MONZ AS.002301) that appeared to be one or two mouls from adulthood was larger (body length = 9.3, carapace length = 3.9) than five of the 11 adult specimens of *P. suterii* examined; based on this and the size of the male, it would be reasonable to expect an adult female *P. keani* to be larger than any *P. suterii* female.
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APPENDIX A: Glossary of technical terms

abdomen — posterior division of the spider body; sometimes referred to as the opisthosoma
bulb — refers to the male pedipalpal organ as a whole
carapace — the hard dorsal covering of the cephalothorax
cephalothorax — anterior division of the spider body; sometimes referred to as the prosoma
chelicerae — first pair of appendages of the cephalothorax, consisting of two segments (the distal segment is called the fang, basal segment is called the paturon)
coxa — first or basal segment of the legs
cymbium — tarsus of the adult male pedipalp
distal — near the apex
dorsal — upper (surface)
epigastric furrow — a transverse groove across the anterior ventral part of the abdomen
epigyne — the sclerotised region of female spiders covering the internal genitalia and located between the book lungs and anterior of the epigastric furrow
embolus — the intromittent structure of the bulb containing the terminal portion of the ejaculatory duct
femur — third segment of the legs and pedipalps
fovea — depression on the thoracic region of the carapace where muscles of sucking stomach are attached internally
haplogyne — the primitive form of spider genitalia where the female has the copulatory openings internally, within the gonopore, and typically lacks a sclerotised epigyne, and the male has relatively simple pedipalps. This form is found in the basal spiders, including Mygalomorphae, Gradungulidae and Haplogyne.

maxilla — (= endite) the expanded lobe of the palpal coxa situated laterally of the labium (plural maxillae)
metatarsus — sixth segment of the legs; absent in the pedipalps
patron — the basal segment of a chelicerae
pedipalp — six-segmented second appendage of the cephalothorax, anterior to legs I
patella — fourth segment of the legs and pedipalp
promarginal — anterior margin
proximal — near the base
retrolateral — on the outer side i.e., the surface nearer to the posterior end of the body
retromarginal — posterior margin
sclerotised — hardened by sclerotin or other substances in the cuticle
seta — a sclerotised hair-like projection arising from the cuticle (plural setae)
sternum — plate on the ventral surface of the cephalothorax between the coxae of the legs
tarsus — last segment of the legs and pedipalp
tibia — fifth segment of legs and pedipalp
trochanter — second segment of the leg and pedipalp
ventral — lower (surface)
APPENDIX B: Collection details of specimens examined. Localities (including coordinates) and dates collected, collectors, and institutions of specimens examined.

**Periegops suterii**

**MC.** 3 ♂, 1 ♀, Riccarton Bush, 43°31.7’S, 172°35.7’E, 9 Apr 1994, A. D. Blest, OMNZ IV35925; 1 ♂, Kennedy Bush Reserve, 43°37.9’S, 172°37.3’E, 22 Apr 2011, C. J. Vink, H. P. & L. J. Hudson Vink, LUNZ 00012715; 1 ♂, Kennedys Bush, 43°37.3’E, 13 Aug 2011, C. J. Vink & H. P. Hudson Vink, LUNZ 00012719; 1 ♂, Rhodes Bush, 43°40’S, 172°37’E, 13 Nov 1915, G. Archey, OMNZ IV35927; 1 ♂, Hay Reserve, 43°42.18’S, 172°53.87’E, 13 Mar 2003, C. J. Vink, LUNZ 00012720; 1 ♂, Kai-tuna Valley, 43°43’S, 172°45’E, 11 Sep 1949, R. R. Forster, OMNZ IV35923; 1 ♂, Montgomery Park Reserve, 43°44.7’S, 172°52.2’E, 5 Nov 2010, C. J. Vink & J. Malumbres-Olarte, LUNZ 00012721; 1 ♂, Montgomery Park Reserve, 43°44.7’S, 172°52.2’E, 5 Nov 2010, C. J. Vink & J. Malumbres-Olarte, LUNZ 00012722; 1 ♂, Montgomery Park Reserve, 43°44.7’S, 172°52.2’E, 5 Nov 2010, C. J. Vink & J. Malumbres-Olarte, LUNZ 00012723; 1 ♂, Montgomery Park Reserve, 43°44.7’S, 172°52.2’E, 5 Nov 2010, C. J. Vink & J. Malumbres-Olarte, LUNZ 00012724; 1 ♂, Montgomery Park Reserve, 43°44.7’S, 172°52.2’E, 5 Nov 2010, C. J. Vink & J. Malumbres-Olarte, LUNZ 00012725; 1 ♂, Panama Rock, 43°44.77’S, 173°02.49’E, 12 Jan 2011, M. H. Bowie, LUNZ 00012726; 1 ♂, Panama Rock, 43°44.7’S, 173°02.49’E, 12 Jan 2011, M. H. Bowie, LUNZ 00012727; 1 ♂, Otepatotu Reserve, 43°44.9’S, 173°00.95’E, 4 Jan 2011, M. H. Bowie, LUNZ 00012728; 1 ♂, Otepatotu Reserve, 43°44.9’S, 173°00.95’E, 12 Jan 2011, M. H. Bowie, LUNZ 00012729; 1 ♂, Otepatotu Reserve, 43°44.9’S, 173°00.95’E, 12 Jan 2011, M. H. Bowie, LUNZ 00012730; 1 ♂, Otepatotu Reserve, 43°44.9’S, 173°00.95’E, 4 Jan 2011, M. H. Bowie, LUNZ 00012731; 1 ♂, Little River, 43°46’S, 172°47’E, 10 Jan 1985, A. C. Harris, OMNZ IV35924; 1 ♂, Ellangowan Reserve, 43°47.89’S, 173°02.08’E, 12 Jan 2011, M. H. Bowie, LUNZ 00012733; 1 ♂, Hinewai Reserve, 43°48.59’S, 173°01.28’E, 11 Oct 2002, C. J. Vink, AMNH ARAMR-000615; 1 ♂, Hinewai Reserve, 43°48.59’S, 173°01.28’E, 13 Oct 2011, M. H. Bowie, LUNZ 00012734; 1 ♂, Hinewai Reserve, 43°48.6’S, 173°01.3’E, 27 Aug 1996, C. J. Vink, LUNZ 00012735; 1 ♂, Akaroa, 43°48.8’S, 172°57.5’E, 16 Oct 1920, G. Archey, OMNZ IV35928; 1 subadult ♂, Fishermans Bay, 43°49.54’S, 173°03.85’E, 12 Dec 2010, M. H. Bowie, LUNZ 00012736.

**Periegops keani**

Fig. 1 Schematic dorsal view, *Periegops suterii*, female (LUNZ 00012720). Actual size on the right.
Fig. 2, 3 *Periegops keani* (holotype): (2) anterior view of chelicerae; (3) ventral view of mouthparts.
Fig. 4, 5 Retrolateral view of leg I: (4) *Periegops suteri* (LUNZ 00012722); (5) *Periegops keani* (holotype).
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Fig. 10, 11 Ventral view of female epigastric area: (10) Periegops suterii, Hay Reserve (LUNZ 00012720); (11) Periegops australia, Mt Goonaneman (QMB S20419).
Fig. 12, 13 Dorsal view of female internal genitalia (p – poreplate; ic – invaginated cup): (12) *Periegops suterii*, Hay Reserve (LUNZ 00012720); (13) *Periegops australia*, Mt Goonaneman (QMB S20419).
Fig. 14 Dorsal view of *Periegops suterii*, male (LUNZ 00012721). Actual size on the right.
Fig. 15 Dorsal view of *Periegops keani*, male (holotype). Actual size on the right.
Fig. 16 Photograph of *Periegops suterii*, female (LUNZ 00012718). Photographer: Bryce McQuillan.
Map 1 Collection localities, *Periegops suterii*. Inset shows localities on and near Banks Peninsula and labelled locations indicate collection sites for DNA specimens.

Map 2 Collection localities, *Periegops keani*. 

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